



HOKKAIDO UNIVERSITY

Title	Dermacentor (Indocentor) auratus Supino 1897: Potential geographic range, and medical and veterinary significance
Author(s)	Teo, Ernest J. M.; Apanaskevich, Dmitry A.; Barker, Stephen C. et al.
Citation	Acta tropica, 254, 107197 https://doi.org/10.1016/j.actatropica.2024.107197
Issue Date	2024-06
Doc URL	https://hdl.handle.net/2115/95868
Rights	© 2024. This manuscript version is made available under the CC-BY-NC-ND 4.0 license https://creativecommons.org/licenses/by-nc-nd/4.0/
Rights(URL)	https://creativecommons.org/licenses/by-nc-nd/4.0/
Type	journal article
File Information	Teo et al., D_auratus_revision_untracke.pdf



Dermacentor (Indocentor) auratus Supino 1897: potential geographic range, and medical and veterinary significance

Ernest J.M. Teo^{1,*}

Dmitry A. Apanaskevich^{2,3}

Stephen C. Barker⁴

Ryo Nakao¹

¹Laboratory of Parasitology, Faculty of Veterinary Medicine, Hokkaido University, Sapporo, Hokkaido 060-0818, Japan

²United States National Tick Collection, The James H. Oliver, Jr. Institute for Coastal Plain Science, Georgia Southern University, Statesboro, GA 30460, USA

³Department of Biology, Georgia Southern University, Statesboro, GA 30460, USA

⁴Department of Parasitology, School of Chemistry & Molecular Biosciences, The University of Queensland, Brisbane, Qld 4072, Australia

*corresponding author

Abstract

Dermacentor (Indocentor) auratus Supino, 1897 occurs in many regions of Southeast Asia and South Asia. In many regions of Southeast Asia and South Asia, targeted tick sampling and subsequent screening of collected *D. auratus* ticks have detected pathogenic bacteria and viruses in *D. auratus*. These disease-causing pathogens that have been detected in *D. auratus* include *Anaplasma*, *Bartonella*, *Borrelia*, *Rickettsia* (including spotted fever group rickettsiae), African swine fever virus, Langkat virus, and Kyasanur forest disease virus. Although *D. auratus* predominantly infests wild pigs, this tick is also an occasional parasite of humans and other animals. Indeed, some 91% of human otoacariasis cases in Sri Lanka were due to infestation by *D. auratus*. With the propensity of this tick to feed on multiple species of hosts, including humans, and the detection of pathogenic bacteria and viruses from this tick, *D. auratus* is a tick of medical, veterinary, and indeed zoonotic concern. The geographic range of this tick, however, is not well known. Therefore, in the present paper, we used the species distribution model, BIOCLIM, to project the potential geographic range of *D. auratus*, which may aid pathogen and tick-vector surveillance. We showed that the potential geographic range of *D. auratus* is far wider than the current geographic distribution of this tick, and that regions in Africa, and in North and South America seem to have suitable climates for *D. auratus*. Interestingly, in Southeast Asia, Borneo and Philippines also have suitable climates for *D. auratus*, but *D. auratus* has not been found in these regions yet despite the apparent close proximity of these regions to Mainland Southeast Asia, where *D. auratus* occurs. We thus hypothesize that the geographic distribution of *D. auratus* is largely dependent on the movement of wild pigs and whether or not these wild pigs are able to overcome dispersal barriers. We also review the potential pathogens and the diseases that may be associated with *D. auratus* and provide an updated host index for this tick.

Key words: Zoonoses, Disease, Public health, Species distribution model, BIOCLIM

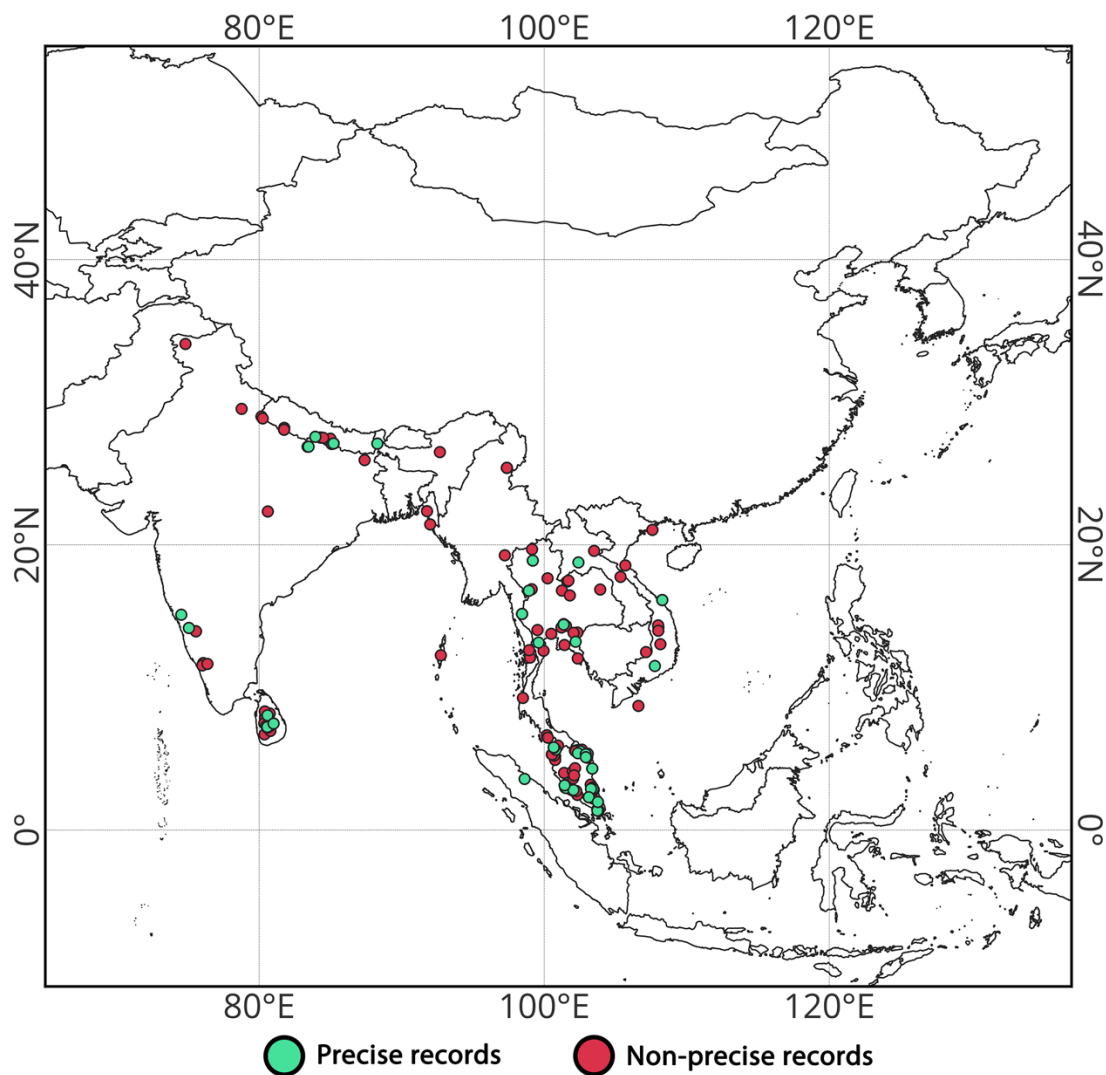


Figure 1. Records of *Dermacentor auratus*. In green, records that we considered to be precise (precision $\leq 5\text{km}$) and in red, records that we considered to be non-precise (precision $> 5\text{km}$). Precise records were used for modelling the distribution of *D. auratus*.

Dermacentor (Indocentor) auratus Supino, 1897, is the most widely distributed *Indocentor* tick, occurring in much of Mainland Southeast Asia and South Asia (Petney and Keirans, 1996) (Fig. 1). Although *D. auratus* is predominantly a parasite of wild pigs (*Sus* spp.), periodic infestation of other animals and humans are common (Hoogstraal and Wassef, 1985; Parola et al., 2003; Liyanaarachchi et al., 2015) (Fig. 2). Moreover, there have been multiple detections of pathogenic bacteria and viruses of importance to human and animal health from *D. auratus*, and thus *D. auratus* is a tick of medical, veterinary, and indeed, zoonotic importance.

As with other ticks from Southeast Asia (Petney et al., 2007), the ecology, the vector competence, and the geographic distribution of *D. auratus* is not well established. This is of particular concern given the importance of *D. auratus* in human and animal health. Therefore, in the present paper, we project the potential geographic range of *D. auratus* with the species distribution model BIOCLIM. Species distribution models allow us to project the potential geographic range of a species when systematic surveys are not feasible and/or when presence data is sparse; such is the case for *D. auratus*. We also review the pathogens and the diseases that are associated with *D. auratus*, a tick that seems to be one of the most important ticks to human and veterinary medicine in parts of Asia.

2.0 Materials & Methods

2.1 Climate data

Mean near-surface air temperature (hereafter, mean temperature), vapor pressure deficit, and precipitation from CHELSA v2.1 (Karger et al., 2017; Brun et al., 2022) were used. We chose these variables because the activity, the abundance, and the geographic distribution and geographic range of ticks seem to be closely related to these climatic variables (Randolph and Storey, 1999; Zemtsova et al., 2016; Teo et al., 2021a; Teo et al., 2021b; Teo et al., 2023). [We use ‘geographic distribution’ to indicate where the tick has been found i.e., presences; whereas ‘geographic range’ describes where the tick may be found spatially i.e., range of suitable climate.] The climatic variables were available at a resolution of 30 arc seconds (< 1km), for each month of the year. Mean temperature and vapor pressure deficit were available as monthly averages whereas precipitation was available as the total precipitation for that month (Karger et al., 2017; Brun et al., 2022). For our analyses, we used these variables from the climatology of 1981-2010 and averaged these variables to obtain a snapshot of the average conditions from 1981-2010.

2.2 Occurrence records of *Dermacentor (Indocentor) auratus*

A Citation Report of publications of *D. auratus* was extracted from the Web of Science Core Collection (<https://www.webofscience.com>) in September 2023 using the search term: ‘*Dermacentor auratus*’, using the default settings: ‘all fields’, and from 1900 to 2023. Additional publications were sought using the search terms: ‘*Indocentor auratus*’, ‘*Dermacentor* Asia’, ‘*Dermacentor* Southeast Asia’, ‘*Dermacentor* India’, ‘ticks Asia’, ‘ticks Southeast Asia’, and ‘ticks India’. Other pertinent publications that had been cited in the

publications from the Citation Report but were not available from the Web of Science Core Collection were also studied. These additional publications are henceforth referred to as 'supplementary publications' for ease of reference. Locality data for *D. auratus* from these publications took one of five forms: (i) precise GPS coordinates; (ii) collection sites plotted on a map; (iii) descriptive localities (e.g. 'tea plantation' in a village); (iv) coarse-scale descriptions (nature reserve/park, village, municipality, or country); and (v) not provided. Georeferencing was done for: (ii), by superimposing the figure provided in the publication on a map of the region; (iii), by studying historical satellite/aerial photographs of the region via Google Earth Pro 7.3.6 and locating the collection site with the provided descriptions; and (iv), by using the approximate center of the nature reserve, nature park, village, municipality, or country. We note that for (iii) and (iv), due to changes in the landscape (e.g. forest-clearing) and/or changes in the names of administrative divisions, some records could not be georeferenced confidently. For (v), when locality data was not provided, we attempted to retrieve information from the provided GenBank accession numbers, if they were available. The retrieved information was then georeferenced. For (i), we checked if the GPS coordinates provided in the respective publication tallied with the descriptors of the location, and if the provided coordinates did not tally with the descriptors provided in the respective publication (e.g. a different coordinate reference system was used), we georeferenced them using one or more of the aforementioned methods.

The georeferenced records were regarded as either 'precise' or 'non-precise'. Precise records were records where the coordinates provided in the respective publications indeed tallied with the location descriptors, records that were georeferenced by superimposing the figures onto a map, or records from regions such as, but not limited to, forests, towns and/or villages that we considered to be small (< 5 km, longest Euclidean distance). Non-precise records on the other hand were records from regions such as, but not limited to, forests, nature reserves, towns, cities and/or countries that we considered to be big (> 5 km). Records we were not confident about because of changes in landscapes and/or names were also regarded as 'non-precise'.

Records prior to the description of Wassef and Hoogstraal (1984) were removed because there was confusion regarding the identity of *D. auratus* prior to their description (Guglielmone et al., 2014). Additionally, records from Taiwan were removed since records from Taiwan need confirmation (Guglielmone et al., 2023). Records from China were also removed as sequences from these records clustered separately from *D. auratus* from India

and from Thailand, even though the latter two clustered together (see Fig. 2 in Wang et al., (2020)). Furthermore, the provided sequences indicated that the records from China were closely related to the records of *D. auratus* from Taiwan that were considered doubtful by Guglielmone et al. (2023) (Supplementary Materials). Hence, like the records from Taiwan, we propose that the records of *D. auratus* from China also require confirmation. We also used the records of and/or field collection sites of *D. auratus* that were available from the U.S. National Tick Collection (USNTC) in our study.

2.3 Potential geographic range of *Dermacentor auratus*

To predict the potential geographic range of *D. auratus*, we used the envelope model BIOCLIM (Nix, 1986). BIOCLIM describes the climatic profile of a species, and identifies locations with potentially suitable environments (Booth et al., 2014). We used BIOCLIM to project the geographic range of *D. auratus* because BIOCLIM only require presences (Nix, 1986; Booth et al., 2014); many of the records of *D. auratus* studied in the present paper were from targeted tick sampling, and therefore do not indicate the full geographic distribution of *D. auratus*. Moreover, envelope models like BIOCLIM are effective at estimating the potential geographic range of a species (Jiménez-Valverde et al., 2008).

Due to the low number of records of *D. auratus* that we considered to have precise locations, and that many of these records were from the same sites, the records were divided into two subequal groups, Subset A and Subset B, by latitude and by longitude for down-stream cross-validation (i.e. geographically structured cross-validation (Radosavljevic and Anderson, 2014)). Because BIOCLIM uses the percentile distribution of values of each variable to indicate the suitability of locations (Nix, 1986), multiple collections (i.e. clustering) from the same site or neighboring sites with similar climates may skew the percentile distribution of values, and subsequently affect the accuracy of the BIOCLIM model (spatial autocorrelation). Therefore, to account for spatial autocorrelation that may arise from clustering, mean temperature, vapor pressure deficit, and precipitation at locations where *D. auratus* were recorded were extracted, and step-wise Moran's *I* statistics were computed for each of the climate variables at Euclidean distances between 0 to 100 km, at an interval of 1 km with *spdep* (Bivand and Wong, 2018). Informed by the calculated Moran's *I* statistic, we thinned each subset by the three climatic variables independently, with *spThin*, and created ten datasets for each climatic variable (Aiello-Lammens et al., 2015). 'Single-variable BIOCLIM models' were generated with *dismo* (Hijmans et al., 2023) for the sixty datasets (2 subsets × 3

variables $\times$ 10 datasets) with the respective climatic variable (e.g. a mean temperature BIOCLIM model was generated with a dataset that was thinned to reduce potential spatial autocorrelation of mean temperature). High (upper limit) and low (lower limit) temperatures were regarded to restrict the geographic range of *D. auratus* whereas only low moisture (high vapor pressure deficit and/or low precipitation), and not high moisture, was regarded to restrict to the geographic range of *D. auratus*. The resulting projections of suitability from the BIOCLIM models generated from the datasets were then averaged by climate variable for Subset A and Subset B. The subsequent six (2 subsets $\times$ 3 variables) projections were then used to generate a ‘three-variable BIOCLIM model’ per subset. This process allowed us to maximize the number of records used for species distribution modelling while reducing potential spatial autocorrelation due to clustering of records of *D. auratus*.

For each subset, records from the same location (i.e. duplicates) were removed; the remaining records from these subsets were used to evaluate the ‘three-variable BIOCLIM model’ generated with records from the other subset (i.e. cross-validation). Sensitivity, the proportion of presence records that were from regions projected to be suitable by the model, and the area under the curve of the receiver operating characteristic (AUC) (Fielding and Bell, 1997), was calculated as a measure of model fit. AUC was calculated using presence and background points with *modEvA* (Márcia Barbosa et al., 2013). Since the models were evaluated with records from a geographically separated set of data, sensitivity and AUC may be interpreted as the model’s transferability to other regions (Radosavljevic and Anderson, 2014).

The ‘three-variable BIOCLIM models’ generated from the two subsets were then averaged to produce a final model and to produce a map of ‘consensus suitability’. Nix (1986), in the context of BIOCLIM, considered environments bound between the 5th and 95th percentile range to indicate ‘core environments’ of a species, and values that fall outside this range to indicate ‘marginal environments’. Although Nix (1986) also considered the 25th to 75th percentile range, no labels were designated to the 25th to 75th percentile range. In the present paper, we considered the minimum value (i.e. 0th percentile) to the 5th percentile, and the 95th percentile to the maximum value (i.e. 100th percentile) to indicate marginal environments; the 5th to the 25th percentile, and the 75th to the 95th percentile to indicate favorable environments; and the 25th to the 75th percentile to indicate optimal environments. By comparing the distribution of values of our variables with the BIOCLIM algorithm implemented in *dismo*

(Hijmans et al., 2023), regions were considered to be unsuitable for *D. auratus* at a suitability of 0, marginal: > 0 , but < 0.14 ; favorable: $0.14 - 0.52$; and optimal: $0.52 - 1$. All analyses were done in R 4.3.1 (R Core Team, 2023), and in QGIS (QGIS Development Team, 2023).

2.4 Disease-causing pathogens associated with *Dermacentor auratus*

Many of the publications we reviewed for the present paper were on the detection and identification of disease-causing pathogens that were associated with *D. auratus*. In the present paper, we summarize these findings. We also present a map of where *D. auratus* have been found with the specific disease-causing pathogens.

3.0 Results & Discussion

3.1 Records of *Dermacentor auratus*

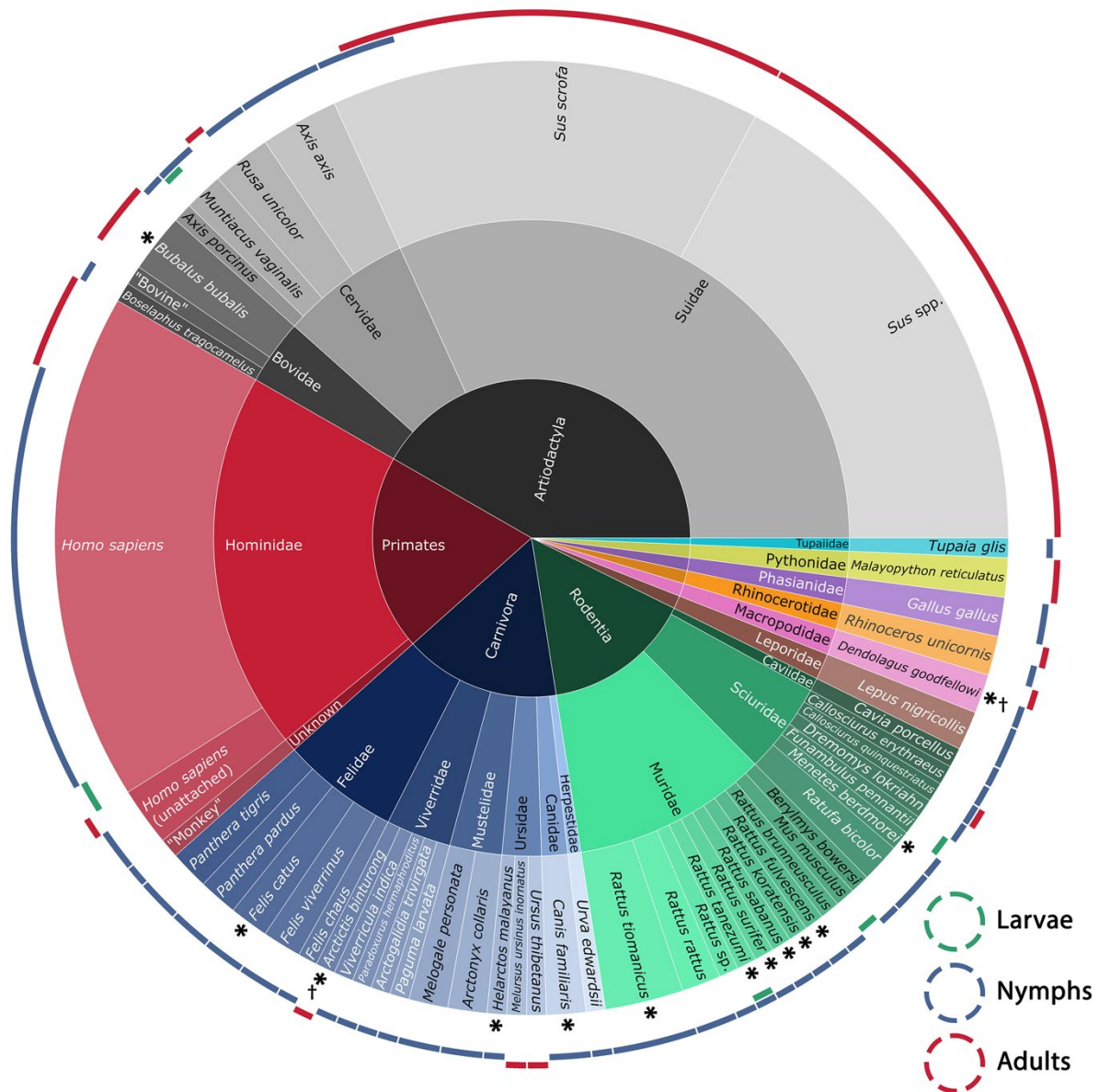
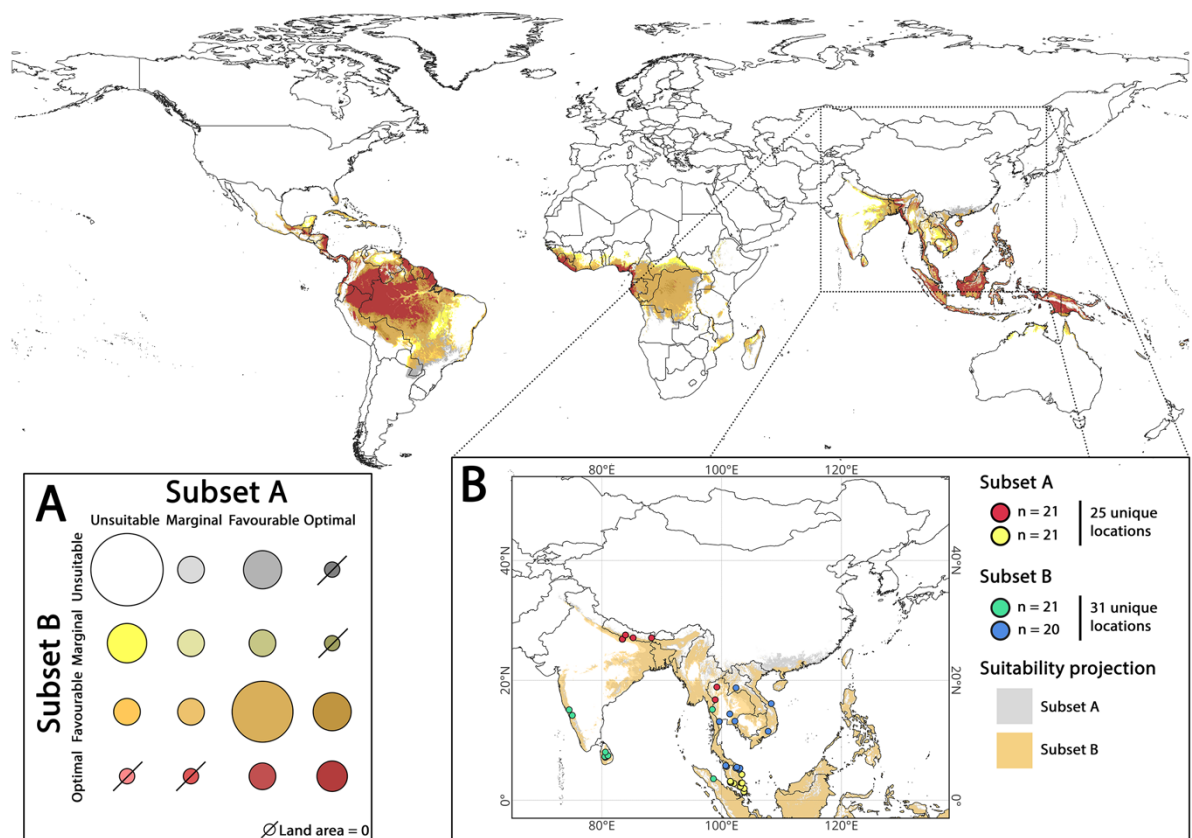


Figure 2. Host records of *Dermacentor auratus* from 151 records that were associated with host species. The three outer disjointed circles indicate the life-stages of *D. auratus* that were associated with the infestation and is shown as a proportion of records that had information of life-stages. Overlaps of the circles indicate host(s) that were infested by multiple life-stages of *D. auratus*. * indicates new host records since Hoogstraal and Wassef (1985) and † indicates that the host was held in captivity. Records of *Sus* spp. were either from *S. scrofa* or *S. barbatus* (Hoogstraal and Wassef, 1985).

Two-hundred and forty-four records of *D. auratus* and/or coordinates of collection sites were retrieved from 28 publications from the Web of Science Core Collection; 13 ‘supplementary publications’; and from the USNTC. These 244 records were from 1935 to 2023 and 25 of these records (10%) that had detected pathogens in *D. auratus* (Tan et al., 1967; Boshell et al., 1968; Sumrandee et al., 2016; Nooroong et al., 2018; Asyikha et al., 2020; Misra et al., 2021; Chaloemthanetphong et al., 2023; Koh et al., 2023). Sixteen of the 244 records were considered doubtful because they were records identified prior to Wassef and Hoogstraal (1984) (n = 8); or were records from Taiwan (Guglielmone et al., 2023) (n = 1) or from China which we considered to be doubtful (n = 7) (Supplementary Materials). Eighty-three of the remaining 228 records were considered to have precise locations (precision ≤ 5 km), and these records were used for our BIOCLIM models (Fig. 1). Also, one hundred and fifty-one of the 228 records were associated with 52 host species; the three most common hosts were: pigs (48/151; 32%), humans (29/151; 19%), and murine rodents (15/151; 10%) (Fig. 2). Thirteen of the 52 host species were new host-parasite associations since Hoogstraal and Wassef (1985) (Fig. 2). Of the three most common hosts, life-stages were recorded for 84% of the records (77/92). Of the 44 records from pigs (Suidae) that had information on the life-stage of the ticks, 42 recorded adults and 3 recorded nymphs (two pigs were infested by adults and by nymphs) (Fig. 2). For humans, of the 21 records that had information on the life-stage of the ticks, 4 recorded adults (3 attached, 1 not attached), 16 recorded nymphs (14 attached, 2 not attached), and 1 recorded an attached larva (Fig. 2). Last, for murine rodents, of the 12 records that had information on the life-stage of the ticks, 11 recorded nymphs, and 2 recorded larvae (one rodent was infested by nymphs and by larvae) (Fig. 2).



239
240 Figure 3. Consensus map of potential geographic range of *Dermacentor auratus* projected by
241 the two BIOCLIM models. Insert A, legend to the main figure and the land area (i.e. number
242 of ~1 km² pixels) by suitability for *D. auratus* by the two BIOCLIM models. Suitable land
243 area was sorted by Jenks natural breaks classification with five breaks. Insert B, geographic
244 distribution of records from Subset A, and from Subset B which were used to generate the
245 two BIOCLIM models. Also, in Subset B, the potential geographic range of *D. auratus* (i.e.
246 range of suitable climates) projected from the two subsets and how well these projections
247 accounted for the records from the other subset.

248
249 Some 88% (22/25) of records of *D. auratus* from Subset A (Fig. 3B) were accounted for by
250 the ‘three-variable BIOCLIM model’ of Subset B. The three records that were not accounted
251 for by the model were between 0.70 to 1.60 km away from the nearest suitable area (mean ±
252 s.d. of 0.94 ± 0.47 km). The AUC for the ‘three-variable BIOCLIM model’ of Subset B was
253 0.85. On the other hand, some 87% (27/31) of records of *D. auratus* from Subset B (Fig. 3B)
254 were accounted for by the BIOCLIM model generated with records from Subset A. The four
255 records that were not accounted for by the model were 0.85 to 9.89 km away from the nearest
256 suitable area (7.31 ± 3.74 km). The AUC for the ‘three-variable BIOCLIM model’ of Subset

A was 0.86. The AUC of the models generated from Subset A and Subset B were 0.86 and 0.85 respectively, which indicated that the models performed better than random ($AUC > 0.5$) (Fielding and Bell, 1997). There was much consensus between the two models: 89% of the land area projected to be suitable (suitability > 0) by the BIOCLIM model of Subset A was also projected to be suitable by the BIOCLIM model of Subset B. The reverse was also true, with a consensus of 85%. Furthermore, all of the regions projected to be optimal by the BIOCLIM model of one subset was projected to either be favorable or optimal by the BIOCLIM model from the other subset (Fig. 3).

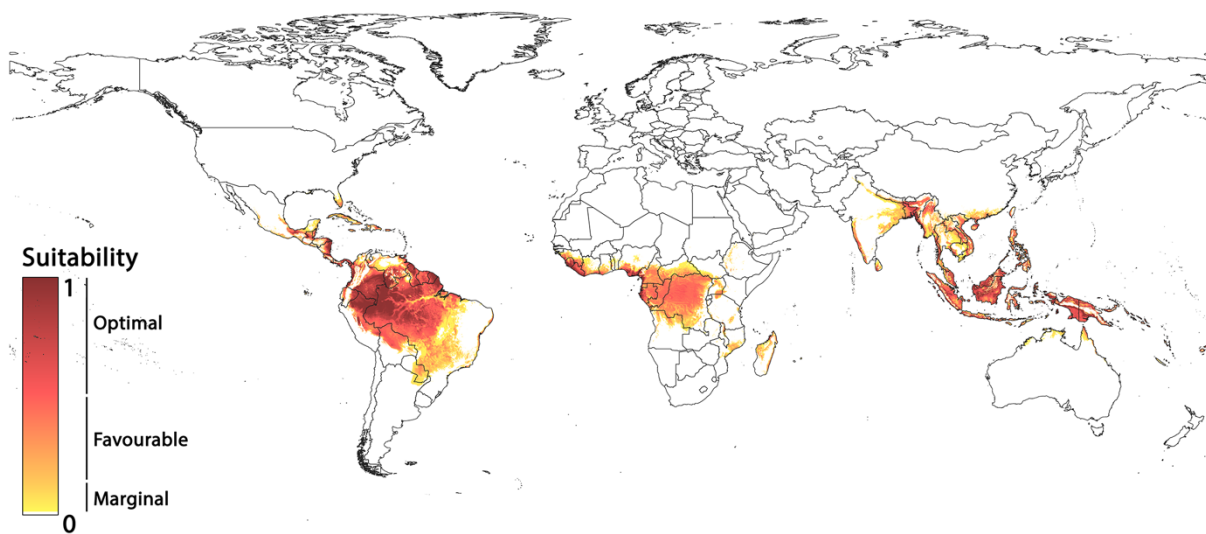


Figure 4. Potential geographic range of *Dermacentor auratus* from the final model. Suitability was averaged from the two BIOCLIM models. White indicates regions which were projected to be unsuitable by both BIOCLIM models (suitability of 0). Regions projected to be suitable (suitability > 0) by at least one BIOCLIM model on the other hand, are shaded, with darker colors indicating higher suitability.

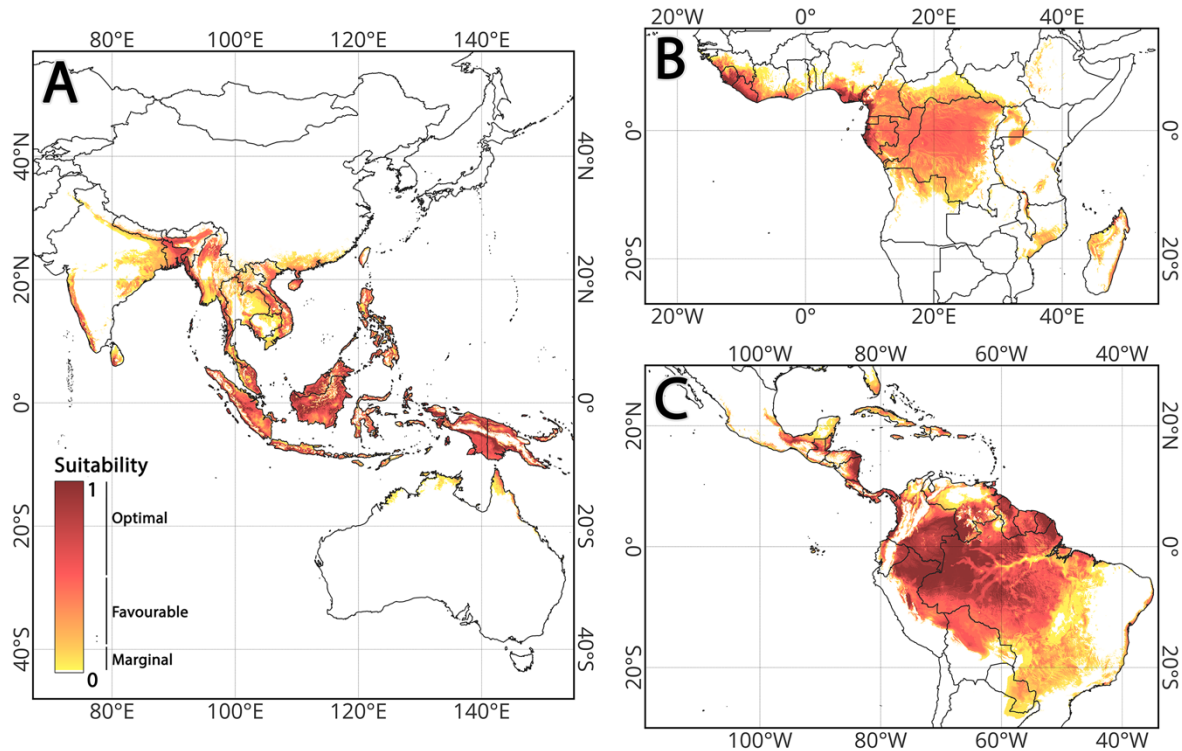


Figure 5. Potential geographic range of *Dermacentor auratus* in three regions. A, parts of Asia and Australia; B, parts of Africa; and C, parts of North and South America.

Although the geographic distribution of *D. auratus* is more-or-less constrained to South East Asia and parts of South Asia (Fig. 1 & 3), the potential geographic range of this tick (i.e. extent of regions with suitable climates) is much greater, with the northern parts of Australia, parts of Africa, parts of North America and parts of South America projected to be climatically suitable for *D. auratus* (Figs. 3, 4 & 5). We note with interest that *D. auratus* has only been recorded on Mainland Southeast Asia and South Asia, and from islands and island countries that are in very close proximity to Mainland Southeast Asia and South Asia (e.g. Singapore, Sri Lanka, and a single record from Sumatra) (Fig. 1). To our knowledge, *D. auratus* has not been recorded from other parts of Sumatra or from the other islands of Indonesia. *D. auratus* has also not been recorded from Borneo, or from Philippines even though these regions were projected to be suitable by our model and even though these regions are close to Mainland Southeast Asia (Figs. 3, 4 & 5). The possible reintroduction of *D. auratus* into the island country of Singapore by wild pigs were discussed by Kwak et al. (2021). Following studies on the movement of wild pigs into Singapore (Lamperty et al., 2023) and because wild pigs seem to be the predominant as well as a very important host in the life-cycle of *D. auratus* (Hoogstraal and Wassef, 1985), we hypothesize that the dispersal

of *D. auratus* is largely dependent on the movement of wild pigs and the ability of wild pigs to overcome dispersal barriers (e.g. water bodies, hills, and mountain ranges). Moreover, in contrast to other *Indocentor* species whose immatures seem to feed chiefly on rodents (Hoogstraal and Wassef, 1985; Wassef and Hoogstraal, 1988; Apanaskevich and Apanaskevich, 2015) and occasionally, on other medium-sized mammals (e.g. Leporidae, Mustelidae, and Viverridae) (Hoogstraal et al., 1986), immature *D. auratus* have been also been recorded on pigs and other large-sized mammals (e.g. Bovidae, Cervidae, Rhinocerotidae, Ursidae, and Felidae: *Panthera* spp.) (Hoogstraal and Wassef, 1985) (Fig. 2). Compared to other *Indocentor* species, the indiscriminate parasitism of large-sized mammals, which have larger home ranges (Swihart et al., 1988), by immature *D. auratus* may explain why *D. auratus* is the most widely distributed *Indocentor* species (Petney and Keirans, 1996).

Although we hypothesize that the dispersal of *D. auratus* is largely dependent on the movement of wild pigs, *D. auratus* have been imported into the United Kingdom and the United States by travelers and on plants (Keirans and Durden, 2001; Gillingham et al., 2020). There is therefore a possibility that, in the absence of ‘wild pig connectivity’ to Mainland Southeast Asia and South Asia, the geographic distribution of *D. auratus* may expand to other regions that were projected to be suitable by our model.

In the present paper, we used the climatology from 1981-2010 to project the potential geographic range of *D. auratus* using the envelope model, BIOCLIM. We note that our use of the 1981-2010 climatology may not have captured, accurately, the climatic conditions from older (1966-1981) and newer (2010-2023) records of *D. auratus*. The climatology of 1981-2010, however, describes the average conditions over 30 years, and should indicate, to some degree or another, the climate that these older and newer records of *D. auratus* could have been exposed to.

As an envelope model, BIOCLIM performs well in projecting the potential geographic range of a species (Jiménez-Valverde et al., 2008; Booth et al., 2014). The realized geographic range of a species is often smaller than the potential geographic range, and thus, it is likely that the actual potential geographic range of *D. auratus* is smaller than projected. On the contrary, because BIOCLIM uses the percentile distribution of each variable to inform suitability, the range of potential suitable area projected by our model may be smaller than the actual potential geographic range of *D. auratus*. This is because BIOCLIM does not account

for possible interactions between variables e.g. the effects of high mean temperatures may be mitigated by high precipitation (Nix, 1986). In the present paper, however, we used climatic variables that relate well to the biology of ticks (Randolph and Storey, 1999). Moreover, using similar envelope modelling approaches, we have previously shown that these climatic variables were able to account for the presences as well as the absences of other species of ticks to a remarkable extent (Teo et al., 2021a; Teo et al., 2021b).

3.3 Medical and veterinary significance of *Dermacentor auratus*

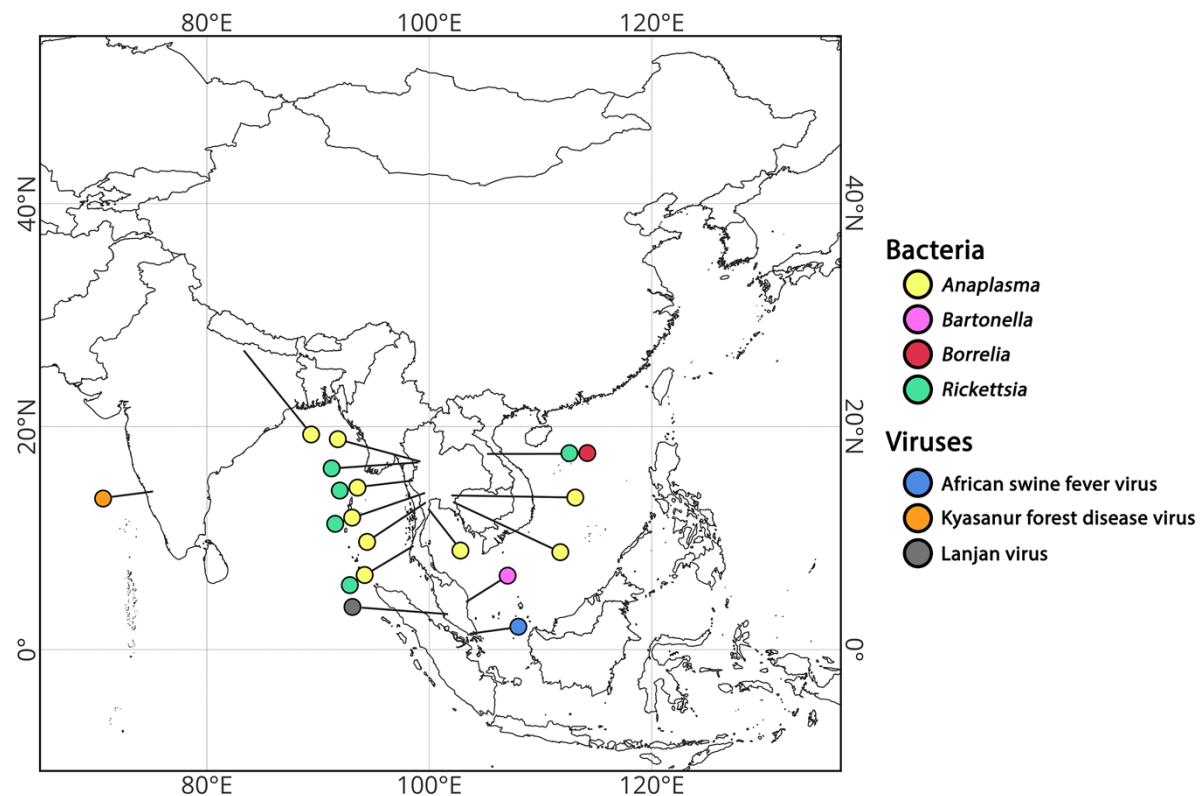


Figure 6. Geographic locations where disease-causing pathogens were detected from *Dermacentor auratus* ticks. Data from Tan et al. (1967); Boshell et al. (1968); Parola et al. (2003); Sumrandee et al. (2016); Taylor et al. (2016); Nooroong et al. (2018); Asyikha et al. (2020); Chaloeanthanetphong et al. (2023); and Koh et al. (2023).

3.3.1 Human infestation

Human infestation by *D. auratus* is normally associated with otoacariasis. Indeed, some 91% of otoacariasis cases in Sri Lanka were due to *D. auratus* and majority of these patients were < 10 years old (Ariyaratne et al., 2016; Bandaranayaka et al., 2021). The most common symptoms associated with otoacariasis is ‘ear fullness’, and tinnitus, with 98-100% and 60-81% of the patients reporting these symptoms respectively (Bandaranayaka et al., 2021).

Other complications of otoacariasis may also include otalgia, secondary bacterial infections, and temporary deafness (Ariyaratne et al., 2016; Bandaranayaka et al., 2021). Analyses of associated risk factors of human infestation by ticks and otoacariasis in Sri Lanka showed that outdoor activities, the presence of wildlife, and the location of the house to be significantly correlated with human infestation by ticks and significantly correlated with otoacariasis (Ariyaratne et al., 2016; Bandaranayaka et al., 2021). Apart from infestation of the ear canal in humans, various feeding life-stages of *D. auratus* has also been recovered from the eyelid as well as other parts of the body from different regions of the world (Kirwan, 1935; Hoogstraal and Wassef, 1985; Ajithkumar et al., 2012; Chitimia-Dobler et al., 2021).

An additional complication of human infestation by *D. auratus* is tick paralysis (Gothe et al., 1979; Hoogstraal and Wassef, 1985). Although the mechanisms of tick paralysis caused by *D. auratus* has not been fully elucidated, a correlation study between otoacariasis and facial nerve palsy has suggested that *Rickettsia* transmitted by the tick could be the causative agent of paralysis associated with feeding *D. auratus* (Kularatne et al., 2018). With high prevalence of human infestation by *D. auratus*, the reason why *D. auratus* may cause paralysis warrants further study.

3.3.2 Bacteria

3.3.2.1 *Rickettsia*

Since Hoogstraal and Wassef (1985), there have been multiple detections of *Rickettsia* spp. from *D. auratus*. Parola et al. (2003) detected *Rickettsia* sp. closely related to *R. bellii* and *Rickettsia* sp. closely related to *R. conorii* subsp. *raoultii*, a spotted fever group rickettsiae (SFGR) from Kanchanaburi province, Thailand (Fig. 6). Similarly, *Rickettsia* sp. that was closely related to *R. conorii* subsp. *raoultii* was also detected in Phang Nga Province, Thailand (Sumrandee et al., 2016) (Fig. 6). From Tak Province, Thailand, *Candidatus Rickettsia laoensis* was detected from questing *D. auratus* (Chaloemthanetphong et al., 2023). Last, there have also been isolations of unidentified *Rickettsia* sp. from Kanchanaburi, Thailand (Nooroong et al., 2018) and from Khammouan Province, Laos from *D. auratus* (Taylor et al., 2016). Despite the numerous isolations of *Rickettsia* and indeed, SFGR, from *D. auratus*, the prevalence of *Rickettsia* spp. seems to be low, at 0 – 27%. We note that like Marchette (1966), none of these studies detected typhus-group *Rickettsia* from *D. auratus*.

Further, a correlation study in Sri Lanka has suggested that SFGR transmitted by ticks could be the causative agent of facial nerve palsy in otoacariasis patients (Kularatne et al., 2018). SFGR, however, was not detected from the single *D. auratus* that was recovered from a patient with facial nerve palsy (Kularatne et al., 2018). SFGR was also not detected in eight *D. auratus* from Sri Lanka in another study (Liyanaarachchi et al., 2016). *D. auratus*, however, is the tick most commonly associated with otoacariasis in Sri Lanka (91%) (Bandaranayaka et al., 2021), where facial nerve palsy associated with otoacariasis is commonplace (Kularatne et al., 2018). The association of *D. auratus* with SFGR in other regions of the world, and the correlation of *D. auratus* with facial nerve palsy associated with SFGR warrants further study since *D. auratus* is an occasional parasite of humans (Fig. 2).

3.3.2.2 *Anaplasma*

There have been multiple detections of *Anaplasma* spp. from *D. auratus*. From Uttar Pradesh, India, *Anaplasma* sp. closely related to *A. centrale* was detected in *D. auratus* that were collected from buffalos (Bovidae) (Misra et al., 2021). From Kanchanaburi province and from Phang Nga Province, Thailand, *Anaplasma* sp. closely related to *A. platys* was detected in *D. auratus* that were collected from dogs and a wild pig respectively (Parola et al., 2003; Sumrandee et al., 2016). Last, Nooroong et al. (2018) detected *Anaplasma* from *D. auratus* collected from vegetation in multiple regions of Thailand. Amongst the detected *Anaplasma*, Nooroong et al. (2018) reported that *Anaplasma* sp. closely related to *A. phagocytophilum* was detected from Prachuap Khiri Khan Province, Thailand (Nooroong et al., 2018). We note with interest that the *Anaplasma* spp. detected in these studies were all closely related to pathogenic *Anaplasma* spp. which are zoonotic, and is of concern (Stuenkel et al., 2013; Arraga-Alvarado et al., 2014).

3.3.2.3 *Bartonella*

A strain of *Bartonella*, closely related to *B. phoceensis* was detected from *D. auratus* parasitizing *Rattus tiomanicus*, the Malayan field rat, from Terengganu, Malaysia (Asyikha et al., 2020). Apart from Malaysia, *B. phoceensis* has also been detected from rodents in Indonesia, Laos, Nepal, and Thailand (Jiyipong et al., 2014), where *D. auratus* occurs. Although *B. phoceensis* has also been detected from non-rodent hosts such as dogs and cats (Tsai et al., 2011; Dybing et al., 2016), *B. phoceensis* is, at the present moment, not considered to be of zoonotic importance (Jiyipong et al., 2014; Dybing et al., 2016).

3.3.2.4 *Borrelia*

There was a single detection of an unidentified *Borrelia* sp. from a nymphal *D. auratus* from Khammouan Province, Laos (Taylor et al., 2016). Similarly, a study in Khao Yai National Park, Thailand, detected *B. theileri*, the causative agent of bovine borreliosis, in unidentified questing larval *Dermacentor* (Takhampunya et al., 2021). Takhampunya et al. (2021), however, did not detect any *Borrelia* from any of the adult *Dermacentor* spp., including *D. auratus*. It is thus not fully known whether or not *D. auratus* is indeed a vector of (i.e. able to transmit) *Borrelia* spp..

3.3.3 Viruses

3.3.3.1 African swine fever virus

African swine fever (ASF) virus (ASFV) (*Asfarviridae*) was detected for the first time from an adult male and an adult female *D. auratus* collected from the carcass of a wild pig (*Sus scrofa*) that was infected with ASFV in Singapore (Koh et al., 2023). It is not known, however, whether or not *D. auratus* is a competent vector of ASFV since both ticks had been attached i.e., whether or not the detected ASFV was from the bloodmeal of the infected wild pig (Koh et al., 2023). We note with interest that ASFV was also detected in *Dermacentor nuttalli*, *Dermacentor niveus*, and *Dermacentor silvarum* in Xinjiang, China (Chen et al., 2019). In the same study, it was shown that *D. niveus* may be considered a reservoir for ASFV since there was transovarial transmission of ASFV; *D. nuttalli* and *D. silvarum* were not tested (Chen et al., 2019). Contrastingly, a study of the replication of ASFV in artificially fed *Dermacentor reticulatus* concluded that although the virus may continue to persist in *D. reticulatus* for weeks, *D. reticulatus* was not considered a reservoir for ASFV because ASFV did not replicate in *D. reticulatus* (de Carvalho Ferreira et al., 2014). It is thus not known whether or not *D. auratus* could be a reservoir of ASFV like *D. niveus*, or whether *D. auratus* is not considered to be a reservoir for ASFV like *D. reticulatus*. The findings from the two studies (de Carvalho Ferreira et al., 2014; Chen et al., 2019) indicate that ASFV may continue to persist in *Dermacentor* ticks. There is thus the possibility that *D. auratus*, especially adult males, may serve as mechanical vectors for ASFV like other hematophagous arthropods (Mellor et al., 1987). Indeed, adult male *Dermacentor* may change their attachment sites even after attachment (Bartosik et al., 2019) which at least to us, indicate that male *Dermacentor* may feed on multiple hosts, resulting in ASFV transmission. On the other hand, although adult female *Dermacentor* will not change their attachment sites if undisturbed (Bartosik et al., 2019), they may attach to another host to feed to repletion if feeding is disturbed (Chen et

al., 2019). Whether or not *D. auratus* does indeed serve as a reservoir of ASFV and/or whether or not *D. auratus* is able to transmit ASFV warrants further study with ongoing outbreaks of ASFV in regions which we projected to be suitable for *D. auratus*.

3.3.3.2 Kyasanur forest disease virus

Kyasanur forest disease (KFD) virus (KFDV) (*Flaviviridae*), the causative agent of KFD or ‘monkey fever’ is a zoonotic pathogen with a human case-fatality rate of 3-10% (Work et al., 1959). *D. auratus* is not considered to be a primary vector of KFDV as there has only been one documented isolation of KFDV from *D. auratus* (Boshell et al., 1968). It was shown in experimental infections, however, that *D. auratus* is a competent vector for KFDV, where transmission of the virus occurred even after moulting to the next life-stage (i.e. transstadial transmission) (Sreenivasan et al., 1979). Furthermore, it was shown that larval and nymphal *D. auratus* are common parasites of rodents (Rajagopalan et al., 1968), and of non-human primates (Trapido et al., 1964) which are the natural hosts for KFDV (Shah et al., 2018). The propensity of immature *D. auratus* to feed on the natural hosts for KFDV; the ability of KFDV to be transmitted transstadially by the tick; the ability of *D. auratus* to transmit the virus to vertebrates; and the propensity of *D. auratus* to feed on humans is thus of concern regarding KFDV transmission to humans by *D. auratus*. We note with interest that the identification of *D. auratus* in KFDV endemic regions (Trapido et al., 1964; Boshell et al., 1968; Rajagopalan et al., 1968; Sreenivasan et al., 1979) precedes the redescription by Wassef and Hoogstraal (1984). *D. auratus*, however, is the only *Dermacentor* known to occur in Karnataka state, India, where these studies were done (Geevarghese et al., 1997).

3.3.3.3 Lanjan virus

Lanjan virus (*Bunyaviridae*) was isolated from nymphs of the ‘*D. auratus*-group’ in Bukit Lanjan, Selangor, Malaysia in 1960 and 1961 (Tan et al., 1967). When inoculated intracerebrally, the virus is pathogenic to suckling and adult mice and results in paralysis and subsequently, death. Although the virus is not pathogenic to adult mice if the mice were inoculated intraperitoneally, it is not known if this route of inoculation is pathogenic to suckling mice because this route of inoculation was not tested (Taylor, 1968). The virus was also not pathogenic to other animals regardless of the route of inoculation (Tan et al., 1967). Via serosurveys in neighbouring forest reserves, antibodies against Lanjan virus were detected from rodents (Muridae) in Ulu Langat Forest Reserve. We note with interest that Hoogstraal and Wassef (1984) indicated that the nymphs found to have Lanjan virus in Tan et

al. (1967) were probably *D. compactus* or *D. atrosignatus* (the latter now referred to as *D. tricuspis* (Apanaskevich et al., 2021)); this cannot be confirmed because the ticks were homogenized in Tan et al. (1967). In their review of the medical significance of *D. auratus*, however, Hoogstraal and Wassef (1985) associated Lanjan virus with *D. auratus*. It is therefore not completely clear whether or not *D. auratus* is a vector of Lanjan virus. We have included Lanjan virus in the present paper as many authors implicate *D. auratus* as the vector of this virus, citing Tan et al. (1967); this will need further clarification.

4.0 Conclusion

Many of the reports of human infestation and/or the detection and isolation of pathogens associated with *D. auratus* were clustered, with most reports of otoacariasis from Sri Lanka, and most detections of disease-causing bacteria from Thailand. Hence, the map of locations where pathogens have been detected from *D. auratus* that we present here may not be comprehensive; that it is likely that other populations of *D. auratus* may also harbor disease-causing pathogens, but they have not been studied in such detail. We note that all the records of *D. auratus* and disease-causing pathogens in the present study are from scientific publications and that each of these publications may or may not have had restrictions pertaining to the sharing of scientific findings prior to the submission of the manuscript. Hence, it is likely that at least some details pertaining to the distribution of *D. auratus* and the pathogens associated with *D. auratus* is under-represented in this paper. The detection of some of these pathogens in *D. auratus* also do not indicate the capacity of *D. auratus* to transmit these pathogens: this warrants further study.

Acknowledgements

EJMT thanks Dr Low Bi Wei and Dr Rayson Lim for stimulating conversations on species distribution modelling. This study was supported, in part, by the Japan Society for the Promotion of Science (JSPS) KAKENHI, Japan (20KK0151, 22H02505 and 23KF0128)

511 **References**

- 512 Aiello-Lammens, M.E., Boria, R.A., Radosavljevic, A., Vilela, B., Anderson, R.P., 2015. spThin:
513 an R package for spatial thinning of species occurrence records for use in ecological niche
514 models. *Ecography* 38, 541-545. doi: 10.1111/ecog.01132
- 515 Ajithkumar, K.G., Ravindran, R., Ghosh, S., 2012. *Dermacentor auratus* Supino, 1897 (Acarina,
516 Ixodidae) reported from Wayanad, Kerala. *Indian J. Med. Res.* 135, 435-436.
- 517 Apanaskevich, D.A., Apanaskevich, M.A., Nooma, W., Ahantarig, A., Trinachartvanit, W.,
518 2021. Reinstatement of *Dermacentor tricuspis* (Schulze, 1933) n. comb., n. stat. (Acari:
519 Ixodidae) as a valid species, synonymization of *D. atrosignatus* Neumann, 1906 and
520 description of a new species from Indonesia, Malaysia and Thailand. *Sys. Parasitol.* 98, 207-
521 230. doi: 10.1007/s11230-021-09972-6
- 522 Apanaskevich, M.A., Apanaskevich, D.A., 2015. Reinstatement of *Dermacentor bellulus*
523 (Acari: Ixodidae) as a valid species previously confused with *D. taiwanensis* and comparison
524 of all parasitic stages. *J. Med. Entomol.* 52, 573-595. doi: 10.1093/jme/tjv034
- 525 Ariyaratne, S., Apanaskevich, D.A., Amarasinghe, P.H., Rajakaruna, R.S., 2016. Diversity and
526 distribution of tick species (Acari: Ixodidae) associated with human otoacariasis and socio-
527 ecological risk factors of tick infestations in Sri Lanka. *Exp. Appl. Acarol.* 70, 99-123. doi:
528 10.1007/s10493-016-0056-z
- 529 Arraga-Alvarado, C.M., Qurollo, B.A., Parra, O.C., Berrueta, M.A., Hegarty, B.C.,
530 Breitschwerdt, E.B., 2014. Case report: molecular evidence of *Anaplasma platys* infection in
531 two women from Venezuela. *Am. J. Trop. Med. Hyg.* 91, 1161-1165. doi: 10.4269/ajtmh.14-
532 0372
- 533 Asyikha, R., Sulaiman, N., Mohd-Taib, F.S., 2020. Detection of *Bartonella* sp. in ticks and their
534 small mammal hosts in mangrove forests of Peninsular Malaysia. *Trop. Biomed* 37, 919-931.
535 doi: 10.47665/tb.37.4.919
- 536 Bandaranayaka, K.O., Kularatne, S.A.M., Rajapakse, R.P.V.J., Abeyesundara, U.B., Rajapaksha,
537 R.M.M.A., Rajakaruna, R.S., 2021. Human otoacariasis in two climatically diverse districts in
538 Sri Lanka: seasonality, risk factors, and case notes. *Acta Parasitol.* 66, 1326-1340. doi:
539 10.1007/s11686-021-00372-w
- 540 Bartosik, K., Buczek, A., Buczek, W., Buczek, A.M., Kulina, D., Koman-Iżko, A., 2019. Host
541 feeding behaviour of *Dermacentor reticulatus* males in relation to the transmission of
542 pathogens. *Ann. Agric. Environ. Med.* 26, 227-230. doi: 10.26444/aaem/105402
- 543 Bivand, R.S., Wong, D.W.S., 2018. Comparing implementations of global and local indicators
544 of spatial association. *TEST* 27, 716-748. doi: 10.1007/s11749-018-0599-x
- 545 Booth, T.H., Nix, H.A., Busby, J.R., Hutchinson, M.F., 2014. bioclim: the first species
546 distribution modelling package, its early applications and relevance to most current MaxEnt
547 studies. *Divers. Distrib.* 20, 1-9. doi: 10.1111/ddi.12144
- 548 Boshell, J., Rajagopalan, P.K., Patil, A.P., Pavri, K.M., 1968. Isolation of Kyasanur forest disease
549 virus from ixodid ticks: 1961-1964. *Indian J. Med. Res.* 56, 541-568.
- 550 Brun, P., Zimmermann, N.E., Hari, C., Pellissier, L., Karger, D.N., 2022. Global climate-related
551 predictors at kilometer resolution for the past and future. *Earth Syst. Sci. Data* 14, 5573-
552 5603. doi: 10.5194/essd-14-5573-2022
- 553 Chaloeanthanetphong, A., Ahantarig, A., Apanaskevich, D.A., Hirunkanokpun, S., Baimai, V.,
554 Trinachartvanit, W., 2023. A novel *Rickettsia*, *Candidatus Rickettsia takensis*, and the first
555 record of *Candidatus Rickettsia laoensis* in *Dermacentor* from Northwestern Thailand. *Sci.*
556 *Rep.* 13, 10044. doi: 10.1038/s41598-023-37206-w

557 Chen, Z., Xu, X., Wang, Y., Bei, J., Jin, X., Dou, W., Ji, H., Duan, Y., Yang, X., Gao, S., 2019. DNA
558 segments of African Swine Fever Virus detected for the first time in hard ticks from sheep
559 and bovines. *Sys. Appl. Acarol.* 24, 180-184.

560 Chitimia-Dobler, L., Schaper, S., Bröker, M., Nava, S., 2021. Long-term itching in a tourist
561 following bite by a nymph of *Dermacentor auratus* (Acari: Ixodidae) in Cambodia. *J. Med.*
562 *Entomol.* 58, 2495-2498. doi: 10.1093/jme/tjab088

563 de Carvalho Ferreira, H.C., Tudela Zúquete, S., Wijnveld, M., Weesendorp, E., Jongejan, F.,
564 Stegeman, A., Loeffen, W.L.A., 2014. No evidence of African swine fever virus replication in
565 hard ticks. *Tick Tick Borne Dis.* 5, 582-589. doi: 10.1016/j.ttbdis.2013.12.012

566 Dybing, N.A., Jacobson, C., Irwin, P., Algar, D., Adams, P.J., 2016. *Bartonella* species identified
567 in rodent and feline hosts from island and mainland Western Australia. *Vector Borne*
568 *Zoonotic Dis.* 16, 238-244. doi: 10.1089/vbz.2015.1902

569 Fielding, A.H., Bell, J.F., 1997. A review of methods for the assessment of prediction errors in
570 conservation presence/absence models. *Environ. Conserv.* 24, 38-49. doi:
571 10.1017/S0376892997000088

572 Geevarghese, G., Fernandes, S., Kulkarni, S.M., 1997. A checklist of Indian ticks (Acari:
573 Ixodoidea). *Indian J. Anim. Sci.* 67, 566-574. doi:

574 Gillingham, E.L., Cull, B., Pietzsch, M.E., Phipps, L.P., Medlock, J.M., Hansford, K., 2020. The
575 unexpected holiday souvenir: the public health risk to UK travellers from ticks acquired
576 overseas. *Int. J. Env. Res. Public Health* 17, 7957. doi: 10.3390/ijerph17217957

577 Gothe, R., Kunze, K., Hoogstraal, H., 1979. The mechanisms of pathogenicity in the tick
578 paralyses. *J. Med. Entomol.* 16, 357-369. doi: 10.1093/jmedent/16.5.357

579 Guglielmone, A.A., Nava, S., Robbins, R.G., 2023. Geographic distribution of the hard ticks
580 (Acari: Ixodida: Ixodidae) of the world by countries and territories. *Zootaxa* 5251, 1-274. doi:
581 10.11646/zootaxa.5251.1.1

582 Guglielmone, A.A., Robbins, R.G., Apanaskevich, D.A., Petney, T.N., Estrada-Peña, A., Horak,
583 I.G., 2014. *The Hard Ticks of the World (Acari: Ixodida: Ixodidae)*, 1st ed. Springer
584 Netherlands : Imprint: Springer, Dordrecht.

585 Hijmans, R.J., Phillips, S., Leathwick, J., Elith, J., 2023. *dismo: species distribution and*
586 *modeling*, 1.3-14 ed.

587 Hoogstraal, H., Wassef, H.Y., 1984. *Dermacentor (Indocentor) compactus* (Acari: Ixodoidea:
588 Ixodidae): wild pigs and other hosts and distribution in Malaysia, Indonesia, and Borneo. *J.*
589 *Med. Entomol.* 21, 174-178. doi: 10.1093/jmedent/21.2.174

590 Hoogstraal, H., Wassef, H.Y., 1985. *Dermacentor (Indocentor) auratus* (Acari: Ixodoidea:
591 Ixodidae): hosts, distribution, and medical importance in tropical Asia. *J. Med. Entomol.* 22,
592 170-177. doi: 10.1093/jmedent/22.2.170

593 Hoogstraal, H., Wassef, H.Y., Santana, F.J., Kuntz, R.E., 1986. *Dermacentor (Indocentor)*
594 *taiwanensis* (Acari: Ixodoidea: Ixodidae): hosts and distribution in Taiwan and southern
595 Japan. *J. Med. Entomol.* 23, 286-288. doi: 10.1093/jmedent/23.3.286

596 Jiménez-Valverde, A., Lobo, J.M., Hortal, J., 2008. Not as good as they seem: the importance
597 of concepts in species distribution modelling. *Divers. Distrib.* 14, 885-890. doi:
598 10.1111/j.1472-4642.2008.00496.x

599 Jiyipong, T., Jittapalapong, S., Morand, S., Rolain, J.-M., 2014. *Bartonella* species in small
600 mammals and their potential vectors in Asia. *Asian Pac. J. Trop. Biomed.* 4, 757-767. doi:
601 10.12980/APJTB.4.2014C742

602 Karger, D.N., Conrad, O., Böhner, J., Kawohl, T., Kreft, H., Soria-Auza, R.W., Zimmermann, N.E.,
 603 Linder, H.P., Kessler, M., 2017. Climatologies at high resolution for the earth's land surface
 604 areas. *Sci. Data* 4, 170122. doi: 10.1038/sdata.2017.122
 605 Keirans, J.E., Durden, L.A., 2001. Invasion: exotic ticks (Acari: Argasidae, Ixodidae) imported
 606 into the United States. A review and new records. *J. Med. Entomol.* 38, 850-861. doi:
 607 10.1603/0022-2585-38.6.850
 608 Kirwan, E.O., 1935. A tick on the upper eye-lid (*Dermacentor auratus* nymph). *Br. J.*
 609 *Ophthalmol.* 19, 659-661. doi: 10.1136/bjo.19.12.659
 610 Koh, E., Tan, A.K.S., Yeo, D., Lau, C., Tan, L.Y., Ng, O.W., Ong, J., Chong, S., Toh, S., Chen, J.,
 611 Wong, W.K., Tan, B.Z.Y., He-Lee, C., Heng, Z.P., Liang, I., Fernandez, C.J., Chang, S.F., Er, K.B.H.,
 612 2023. Detection of African swine fever virus from wild boar, Singapore, 2023. *Emerg. Infect.*
 613 *Dis.* 29. doi: 10.3201/eid2912.230966
 614 Kularatne, S.A.M., Fernando, R., Selvaratnam, S., Narampanawa, C., Weerakoon, K.,
 615 Wickramasinghe, S., Pathirage, M., Weerasinghe, V., Bandara, A., Rajapakse, J., 2018. Intra-
 616 aural tick bite causing unilateral facial nerve palsy in 29 cases over 16 years in Kandy, Sri
 617 Lanka: is rickettsial aetiology possible? *BMC Infect. Dis.* 18, 418. doi: 10.1186/s12879-018-
 618 3338-8
 619 Kwak, M.L., Chavatte, J.-M., Chew, K.L., Lee, B.P.Y.H., 2021. Emergence of the zoonotic tick
 620 *Dermacentor (Indocentor) auratus* Supino, 1897 (Acari: Ixodidae) in Singapore. *Tick Tick*
 621 *Borne Dis.* 12, 101574. doi: 10.1016/j.ttbdis.2020.101574
 622 Lamperty, T., Chiok, W.X., Khoo, M.D.Y., Amir, Z., Baker, N., Chua, M.A.H., Chung, Y.F., Chua,
 623 Y.K., Koh, J.J.-M., Lee, B.P.Y.-H., Lum, S.K.Y., Mendes, C.P., Ngiam, J., ODempsey, A., Png,
 624 K.G.C., Sovie, A.R., Tan, L., Teo, R., Thomas, N., Tianjiao, L., Tze-Ming, B.L., Loo, A.H.B.,
 625 Wardle, D.A., Luskin, M.S., 2023. Rewilding in Southeast Asia: Singapore as a case study.
 626 *Conserv. Sci. Prac.* 5, e12899. doi: 10.1111/csp2.12899
 627 Liyanaarachchi, D.R., Rajakaruna, R.S., Dikkumbura, A.W., Rajapakse, R.P.V.J., 2015. Ticks
 628 infesting wild and domestic animals and humans of Sri Lanka with new host records. *Acta*
 629 *Trop.* 142, 64-70. doi: <https://doi.org/10.1016/j.actatropica.2014.11.001>
 630 Liyanaarachchi, D.R., Rajakaruna, R.S., Rajapakse, R.P.V.J., 2016. Spotted fever group
 631 rickettsia in ticks infesting humans, wild and domesticated animals of Sri Lanka: one health
 632 approach. *Ceylon J. Sci.* 44, 67. doi: 10.4038/cjsbs.v44i2.7351
 633 Marchette, N.J., 1966. Rickettsioses (tick typhus, Q-fever, urban typhus) in Malaya. *J. Med.*
 634 *Entomol.* 2, 339-371. doi: 10.1093/jmedent/2.4.339
 635 Márcia Barbosa, A., Real, R., Muñoz, A.-R., Brown, J.A., 2013. New measures for assessing
 636 model equilibrium and prediction mismatch in species distribution models. *Divers. Distrib.*
 637 19, 1333-1338. doi: 10.1111/ddi.12100
 638 Mellor, P.S., Kitching, R.P., Wilkinson, P.J., 1987. Mechanical transmission of capripox virus
 639 and African swine fever virus by *Stomoxys calcitrans*. *Res. Vet. Sci.* 43, 109-112.
 640 Misra, B.R., Kumar, N., Kant, R., Deval, H., Singh, R., Pandey, A.K., Behera, S.P., Bondre, V.P.,
 641 2021. Abundance of ticks (Acari: Ixodidae) and presence of *Rickettsia* and *Anaplasma* in ticks
 642 infesting domestic animals from Northern India. *J. Med. Entomol.* 58, 1370-1375. doi:
 643 10.1093/jme/tjaa296
 644 Nix, H.A., 1986. A biogeographic analysis of Australian elapid snakes. *Atlas of elapid snakes*
 645 *of Australia*. Bureau of Flora and Fauna, Canberra.
 646 Nooroong, P., Trinachartvanit, W., Baimai, V., Ahantarig, A., 2018. Phylogenetic studies of
 647 bacteria (*Rickettsia*, *Coxiella*, and *Anaplasma*) in *Amblyomma* and *Dermacentor* ticks in

648 Thailand and their co-infection. Tick Tick Borne Dis. 9, 963-971. doi:
 649 10.1016/j.ttbdis.2018.03.027
 650 Parola, P., Cornet, J.P., Sanogo, Y.O., Miller, R.S., Thien, H.V., Gonzalez, J.P., Raoult, D., Telford,
 651 I.S., Wongsrichanalai, C., 2003. Detection of *Ehrlichia* spp., *Anaplasma* spp., *Rickettsia* spp.,
 652 and other eubacteria in ticks from the Thai-Myanmar border and Vietnam. J. Clin. Microbiol.
 653 41, 1600-1608. doi: 10.1128/jcm.41.4.1600-1608.2003
 654 Petney, T., Keirans, J.E., 1996. Ticks of the genera *Boophilus*, *Dermacentor*, *Nosomma*, and
 655 *Rhipicephalus* (Acari: Ixodidae) in South-east Asia. Trop. Biomed. 13, 73-84.
 656 Petney, T.N., Kolonin, G.V., Robbins, R.G., 2007. Southeast Asian ticks (Acari: Ixodida): a
 657 historical perspective. Parasitol. Res. 101 Suppl 2, S201-205. doi: 10.1007/s00436-007-0687-
 658 4
 659 QGIS Development Team, 2023. QGIS Geographic Information System.
 660 R Core Team, 2023. R: a language and environment for statistical computing. R Foundation
 661 for Statistical Computing, Vienna, Austria.
 662 Radosavljevic, A., Anderson, R.P., 2014. Making better Maxent models of species
 663 distributions: complexity, overfitting and evaluation. J. Biogeogr. 41, 629-643. doi:
 664 10.1111/jbi.12227
 665 Rajagopalan, P.K., Patil, A.P., Boshell, J., 1968. Ixodid ticks on their mammalian hosts in the
 666 Kyasanur Forest disease area of Mysore State, India, 1961-64. Indian J. Med. Res. 56, 510-
 667 525. doi:
 668 Randolph, S.E., Storey, K., 1999. Impact of microclimate on immature tick-rodent host
 669 interactions (Acari: Ixodidae): implications for parasite transmission. J. Med. Entomol. 36,
 670 741-748. doi: 10.1093/jmedent/36.6.741
 671 Shah, S.Z., Jabbar, B., Ahmed, N., Rehman, A., Nasir, H., Nadeem, S., Jabbar, I., Rahman, Z.U.,
 672 Azam, S., 2018. Epidemiology, pathogenesis, and control of a tick-borne disease- Kyasanur
 673 forest disease: current status and future directions. Front. Cell. Infect. Microbiol. 8, 149. doi:
 674 10.3389/fcimb.2018.00149
 675 Sreenivasan, M.A., Bhat, H.R., Naik, S.V., 1979. Experimental transmission of Kyasanur forest
 676 disease virus by *Dermacentor auratus* Supino. Indian J. Med. Res. 69, 701-707.
 677 Stuen, S., Granquist, E., Silaghi, C., 2013. *Anaplasma phagocytophilum*—a widespread multi-
 678 host pathogen with highly adaptive strategies. Front. Cell. Infect. Microbiol. 3. doi:
 679 10.3389/fcimb.2013.00031
 680 Sumrandee, C., Baimai, V., Trinachartvanit, W., Ahantrag, A., 2016. Molecular detection of
 681 *Rickettsia*, *Anaplasma*, *Coxiella* and *Francisella* bacteria in ticks collected from Artiodactyla in
 682 Thailand. Tick Tick Borne Dis. 7, 678-689. doi: 10.1016/j.ttbdis.2016.02.015
 683 Swihart, R.K., Slade, N.A., Bergstrom, B.J., 1988. Relating body size to the rate of home range
 684 use in mammals. Ecology 69, 393-399. doi: 10.2307/1940437
 685 Takhampunya, R., Sakolvaree, J., Chanarat, N., Youngdech, N., Phonjatturas, K.,
 686 Promsathaporn, S., Tippayachai, B., Tachavarong, W., Srinoppawan, K., Poole-Smith, B.K.,
 687 McCardle, P.W., Chaorattanakawee, S., 2021. The bacterial community in questing ticks from
 688 Khao Yai National Park in Thailand. Front. Vet. Sci. 8. doi: 10.3389/fvets.2021.764763
 689 Tan, D.S.K., Smith, C.E.G., McMahon, D.A., Bowen, E.T.W., 1967. Lanjan Virus, a new agent
 690 isolated from *Dermacentor auratus* in Malaya. Nature 214, 1154-1155. doi:
 691 10.1038/2141154a0
 692 Taylor, A.J., Vongphayloth, K., Vongsouvath, M., Grandadam, M., Brey, P.T., Newton, P.N.,
 693 Sutherland, I.W., Dittrich, S., 2016. Large-scale survey for tickborne bacteria, Khammouan
 694 Province, Laos. Emerg. Infect. Dis. 22, 1635-1639. doi: 10.3201/eid2209.151969

Taylor, R.M., 1968. Catalogue of arthropod-borne viruses of the world: a collection of data on registered arthropod-borne animal viruses, First ed. U.S. Department of Health, Education, and Welfare, Public Health Service, Bethesda, Maryland.

Teo, E.J.M., Hailu, S., Kelava, S., Zalucki, M.P., Furlong, M.J., Nakao, R., Barker, D., Barker, S.C., 2021a. Climatic requirements of the southern paralysis tick, *Ixodes cornuatus*, with a consideration of its host, *Vombatus ursinus*, and the possible geographic range of the tick up to 2090. Tick Tick Borne Dis. 12, 101758. doi: 10.1016/j.ttbdis.2021.101758

Teo, E.J.M., Russell, H., Lambert, T., Webster, R., Yappa, A., McDonagh, P., Harper, G., Barker, D., Barker, S.C., 2023. The weather determines the number of cases of tick paralysis in dogs and cats in eastern Australia, caused by *Ixodes holocyclus*, the eastern paralysis tick. Aust. Vet. J. doi: 10.1111/avj.13289

Teo, E.J.M., Vial, M.N., Hailu, S., Kelava, S., Zalucki, M.P., Furlong, M.J., Barker, D., Barker, S.C., 2021b. Climatic requirements of the eastern paralysis tick, *Ixodes holocyclus*, with a consideration of its possible geographic range up to 2090. Int. J. Parasitol. 51, 241-249. doi: 10.1016/j.ijpara.2020.08.011

Trapido, H., Goverdhan, M.K., Rajagopalan, P.K., Rebello, M.J., 1964. Ticks ectoparasitic on monkeys in the Kyasanur forest disease area of Shimoga district, Mysore state, India. Am. J. Trop. Med. Hyg. 13, 763-772. doi: 10.4269/ajtmh.1964.13.763

Tsai, Y.L., Lin, C.C., Chomel, B.B., Chuang, S.T., Tsai, K.H., Wu, W.J., Huang, C.G., Yu, J.C., Sung, M.H., Kass, P.H., Chang, C.C., 2011. *Bartonella* infection in shelter cats and dogs and their ectoparasites. Vector Borne Zoonotic Dis. 11, 1023-1030. doi: 10.1089/vbz.2010.0085

Wang, X., Sun, X., Sun, Y., Chen, K., Zhang, K., Xu, W., Fan, K., Lin, W., Chen, T., Lin, X., Lin, K., Chiu, H.-c., Huang, C., 2020. Identification and molecular analysis of Ixodid ticks (Acari: Ixodidae) infesting wild boars (*Sus scrofa*) and tick-borne pathogens at the Meihua mountain of southwestern Fujian, China. Vet. Parasitol.: Reg. Stud. Rep. 22, 100492. doi: 10.1016/j.vprsr.2020.100492

Wassef, H.Y., Hoogstraal, H., 1984. *Dermacentor (Indocentor) auratus* (Acari: Ixodoidea: Ixodidae): identity of male and female. J. Med. Entomol. 21, 169-173. doi: 10.1093/jmedent/21.2.169

Wassef, H.Y., Hoogstraal, H., 1988. *Dermacentor (Indocentor) steini* (Acari: Ixodoidea: Ixodidae): hosts, distribution in the Malay Peninsula, Indonesia, Borneo, Thailand, and Philippines, and New Guinea. J. Med. Entomol. 25, 315-320. doi: 10.1093/jmedent/25.5.315

Work, T.H., Roderiguez, F.R., Bhatt, P.N., 1959. Virological epidemiology of the 1958 epidemic of Kyasanur Forest disease. Am. J. Public Health Nations Health 49, 869-874. doi: 10.2105/ajph.49.7.869

Zemtsova, G.E., Apanaskevich, D.A., Reeves, W.K., Hahn, M., Snellgrove, A., Levin, M.L., 2016. Phylogeography of *Rhipicephalus sanguineus* sensu lato and its relationships with climatic factors. Exp. Appl. Acarol. 69, 191-203. doi: 10.1007/s10493-016-0035-4