



HOKKAIDO UNIVERSITY

Title	Review : Nicotinic acetylcholine receptors to regulate important brain activity : what occurs at the molecular level?
Author(s)	Nara, Shigetoshi; Yamaguti, Yutaka; Tsuda, Ichiro
Citation	Cognitive Neurodynamics, 18(3), 769-774 https://doi.org/10.1007/s11571-023-09975-7
Issue Date	2023-05-19
Doc URL	https://hdl.handle.net/2115/98680
Rights	© The Author(s) 2023
Rights(URL)	https://creativecommons.org/licenses/by/4.0/
Type	journal article
File Information	Cognitive Neurodynamics_18_2024.pdf





Review: Nicotinic acetylcholine receptors to regulate important brain activity—what occurs at the molecular level?

Shigetoshi Nara¹ · Yutaka Yamaguti² · Ichiro Tsuda³

Received: 15 December 2022 / Revised: 2 April 2023 / Accepted: 19 April 2023
© The Author(s) 2023

Abstract

Herein, we briefly review the role of nicotinic acetylcholine receptors in regulating important brain activity by controlled release of acetylcholine from subcortical neuron groups, focusing on a microscopic viewpoint and considering the non-linear dynamics of biological macromolecules associated with neuron activity and how they give rise to advanced brain functions of brain.

Keywords Neuromodulator · Nicotinic · Acetylcholine · Receptors · Brain activity

Introduction

It is well known that neuromodulators play very important roles in advanced functions of brain. Acetylcholine (ACh) is not a highly complex molecule but a rather simple one, as shown in Fig. 1 and it works as a functional neuro-modulators. Furthermore, it is related to the various advanced functions of brain, such as perception, learning, memory, and attention, etc. [1–3]

This paper is initially prepared as a short summary for a presentation at the International Congress on Cognitive Neurodynamics (ICCN) 2021, postponed due to COVID-19 and held in May 2022 in the form of an online conference. After the conference, considerably greater detail was added, and this review-paper version was completed.

✉ Shigetoshi Nara
nara@ec.okayama-u.ac.jp

Yutaka Yamaguti
y-yamaguchi@fit.ac.jp

Ichiro Tsuda
tsuda@isc.chubu.ac.jp

¹ Graduate School of Natural Science and Technology, Okayama University, 3-1-1 Tushima-naka, Kita-ku, Okayama 700-8530, Japan

² Faculty of Information Engineering, Fukuoka Institute of Technology, 3-30-1 Wajiro-higashi, Higashi-ku, Fukuoka 811-0295, Japan

³ Chubu University Academy of Emerging Sciences/Center for Mathematical Science and Artificial Intelligence, Chubu University, Aichi 487-8501, Japan

In the human body, particularly in the brain, acetylcholine is released by neural cells called “cholinergic cells” that exists in subcortical regions. There are 8 cholinergic cell groups in the brain that release ACh; these groups are designated Ch1 ~ Ch8. Regarding visual perception processing, a particularly important cell group is the nucleus basalis of Meynert (nbM) in the basal fore-brain; its long projecting axons release controlled amounts of acetylcholine to very wide area of neocortical regions in the frontal, parietal, temporal, and even occipital lobes (see Fig. 2).

The reason why acetylcholine has such strong influence on various advanced functions is that, once ACh is weakly adsorbed at specific sites on cell-membrane-embedded biological macromolecules called “receptors”, it activates various signal transduction pathways inside the cell [4]. The specific receptors that respond to ACh are known as “acetylcholine receptors” (AChRs hereafter). It is known that there is a wide variety of AChRs, as briefly introduced below.

AChRs are classified into two major types: the nicotinic Acetylcholine Receptor (nAChR) [5] and the muscarinic Acetylcholine Receptor (mAChR) [6]. In the present paper, we will confine ourselves to nAChR as a matter of convenience, given the limited space available. nAChRs are pentamers consisting of five subunits, each of which is a trans-membrane proteins; the total molecular weight of the five subunits is about 250,000 ~ 350,000, and each of the five subunits is a 4-transmembrane-domain protein. Typical experimental observations and their computer

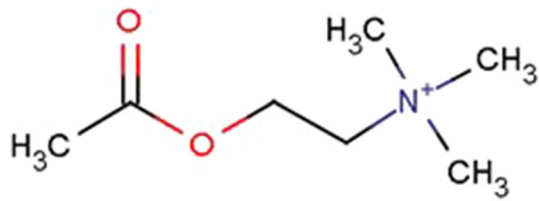


Fig. 1 Acetylcholine molecule. The molecular weight is 146

reconstructions are shown in Fig. 3 [7–9]. To date, despite a long history of investigation into the pertinent mechanisms of acetylcholine receptors, we do not yet have fully clarified the roles of AChRs in advanced brain functions [1].

Phenomena at the microscopic level

On cytoplasmic membrane of neural cells, there are many transmembrane proteins to mediate signals transmission. In such proteins, our concern in this paper is nAChRs introduced in Introduction.

nAChRs belong to a group of proteins called “ion channel proteins”. One of the reasons why nAChRs are of particular interest to the research community, including our team, is that they *regulate neuron firing* by switching their functional state, such as paying attention, memorizing, wakefulness, etc., in a controlled manner to allow sodium ions to pass from the outside to the inside of cell through tiny hollow channel, the diameter of which is on a sub-nanometer scale; the channel has the gate to take either

“the open state” or “the closed state”. The most important point is that the both states are controlled by binding or unbinding of acetylcholine to specific sites on receptors (see Figs. 4 & 5) [10–13]. The papers on this topic is too extensive to discuss within the space limitations here. Readers can familiarize themselves with a large body of experimental data and numerous in-depth considerations through the excellent book written by Changeux & Edelman [14].

Molecular structure and problems

It is known that nAChRs are pentameric in structure and constitute a quite large family depending on different combinations of various kinds of subunits, which are classified into five types according to their amino-acid-sequences: α , β , γ , δ , and ϵ . Of these, the first two have intra-subunit variations, i.e. $\alpha 1 \sim \alpha 10$ and $\beta 1 \sim \beta 4$. The others, γ , δ , and ϵ , do not have distinct subtypes [15–17]. Depending on the combination of subunits in the pentamer, permeability of ion are different under acetylcholine binding, and they are regarded as important factors to regulate advanced functions of brain [1].

Now, let us summarize the important problems regarding these nAChRs. First, from a microscopic viewpoint and nonlinear dynamics of molecular structure, the following problems exist:

- (1) The relaxation time problem, i.e. molecular weights of the receptors are distributed over 250,000 ~ 350,000, while the molecular weight of

Fig. 2 The configuration of the 8 cholinergic cell groups (Ch1 ~ Ch8) in the brain and their projecting axons that release ACh in selected areas in the brain (from <https://kanri.nkdesk.com/>). TH: thalamus, IP: interpeduncular nucleus, HP: hippocampus. It should be noted that the anatomical positions of Hippocampus (HP) and of IP shown in this website should be located a little higher and lower than the described ones, respectively

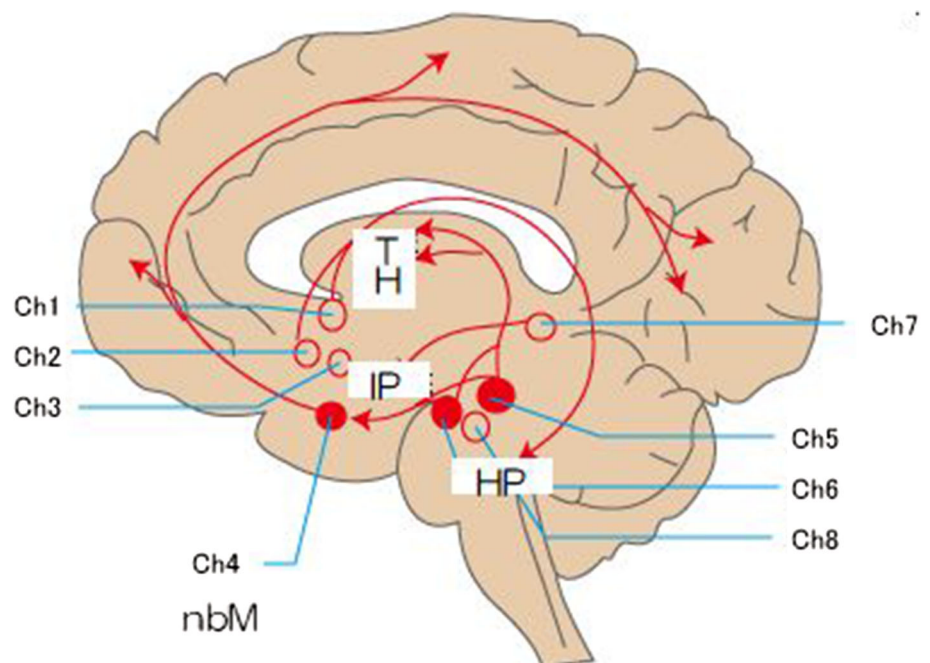


Fig. 3 Typical molecular structure of nAChR described in the following papers. **a** [7] **b** [8]. The estimated largest diameter of the cylindrical molecule is approximately $\sim 10\text{nm}$ or more

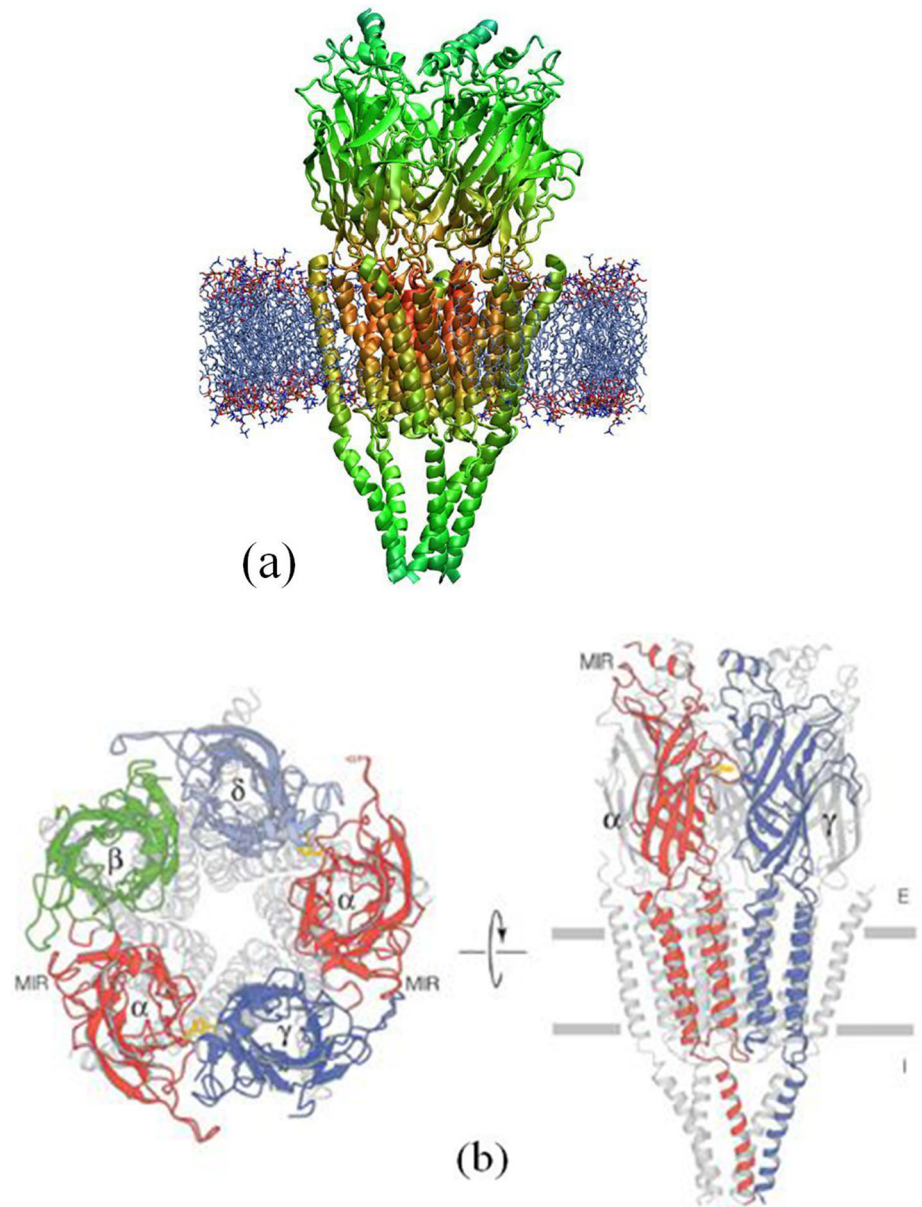
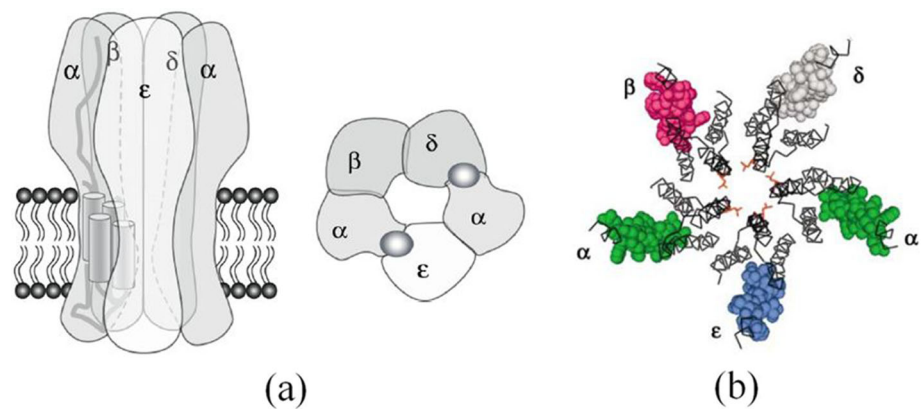


Fig. 4 Configuration of subunits and intra subunit structures to form ion-channel as reported by **a** [12], **b** [13]. The two binding sites of ACh are shown in **a** by the dense grey circle. In **b** at the center of pentagon, there are the five M2- α -helices to form ion-channel



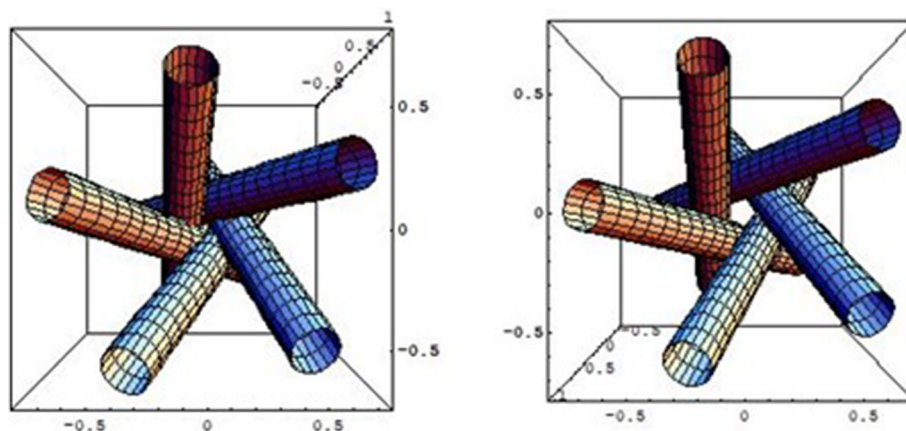


Fig. 5 Schematically described pentagonal configuration consisting of M2- α -helix in five subunits of nAChR drawn by one of the authors (S.N.) with use of Mathematica. The left is closed state and the right is open state of ion-channel. In the former, the upper pentagon formed

by the top of five cylinders and the lower pentagon formed by the bottom of five cylinders are aligned in symmetric configuration. In the latter, the upper pentagon and the lower pentagon are rotated slightly, such that a narrow gap opens

acetylcholine is only 146 and is three or more order of magnitude less than receptors. Thus, binding of acetylcholine to receptors is regarded as a very small physicochemical impact and, in conventional viewpoint of thermal fluctuation based on statistical mechanics, the influence may be relaxed within, at most, nanosecond order. However, after a delay on the order of microseconds following binding, a conformation change in the pentagonal configuration of subunits occurs, and the ion channel at the center of pentagon opens.

- (2) The spatial coherency problem, i.e., the binding position of acetylcholine is far from the position of ion channel, more than $\sim 10\text{nm}$, a distance that seems to be too far to keep dynamically coherent signal transmission in conventional viewpoint of statistical physics in equilibrium systems.
- (3) The energy source problem, i.e., the conformation change associated with ion channel opening is a slight but meaningful distortion and shifting of five M2- α -helices belonging to each subunit. Thermal fluctuation energy or physicochemical energy (adsorbing energy) of ACh is highly probably not enough to cause such movement of five helices.

These are major issues not only in biophysics, biochemistry, and molecular biology [18], but also in nonlinear & non-equilibrium dynamics of systems with huge but finite degrees of freedom related with ‘fluctuation theorem’ that has been extensively discussed since 1993 [19].

The second problem is the complex, entangled relation between releasing control of ACh by cholinergic cell groups in the brain, spatial distribution of nAChRs on neural cells in the brain, and concentration control of acetylcholinesterase (degrading enzyme of ACh). These

three factors govern the total activity of neural networks in the brain and their complex hierarchical regulatory structures from microscopic scale to macroscopic scale are almost out of consideration to deal with the conventional methods.

Extremely rough estimation with respect to the number of neural cells and synaptic connections in human brain may give $\approx 10^{12}$ and $\approx 10^{18}$, respectively. However, the number of synaptic boutons per dendrite are too inhomogeneous to estimate even in averaged sense, although it is said to be $\approx 10^2 \sim 10^4$. While the number of receptors per synaptic bouton is also difficult to estimate, it is said to be $\approx 10^3 \sim 10^5$ but includes great uncertainty. Therefore, one of the important but quite difficult questions is by what mechanism such numerous degrees of freedom are controlled via neural networks to realize many advanced functions. In particular, the most important point is what and how spatiotemporal concentration of ACh regulates advanced functions of brain via nAChRs, say perceiving, learning, memorizing, attentions, etc. [1] A large number of papers have been published so far, however we are not yet able to have clarified understanding [21].

Now, the following is a summary of problems,

- (1) The difficulty in accurate evaluations of the causal links between the spatiotemporal concentration of acetylcholine in the brain and a quantitative evaluation of the performance of advanced functioning (e.g., attention, conscious awareness) and mental diseases (e.g., DLB, PD, AD, etc.) [20, 22].
- (2) The technological problem of measuring the spatial distribution of nAChRs with accurate resolution depending on subunit combinations in nAChRs. [12, 23].

- (3) The technological problem of measuring the spatiotemporal distribution of acetylcholinesterase (degrading enzyme of ACh) that gives strong influences on local concentration of ACh [24]. Unfortunately, most papers are concerned not to measure the data of acetylcholinesterase but to investigate the effects of acetylcholinesterase inhibitors on diseased patients (AD, LBD, etc.). [25].

Concluding remarks and issues

Generally speaking, when we have concerns regarding macroscopic behaviors or motions of animals including humans, then experimental observation and theoretical consideration based on contracting the degrees of freedom to ‘small number of macroscopic variables’ enable us to construct phenomenological theory, e.g. non-equilibrium phase transition or information self-organization [26]. In contrast, if neural activity on *mesoscopic scale* are concerned, then neuro-physiological measurements and theoretical consideration are useful for investigating brain functioning [27], although we have to face a difficulty of ‘big data’ obtained by physiological measurements, e.g. EEG, MEG, ECoG, MRI (fMRI, DTI-MR), SPECT, PET, TMS, etc., all of data appear non-stationary (even chaotic) in wide spatiotemporal scale. However, in the both (macroscopic and mesoscopic) approaches, neuromodulators are out of measurements and consideration.

When one wishes to take the effects of neuromodulators into account, there are few options to observe them in humans because ethical principles forbid arbitrary medical treatments. At most, mild effects by pharmacological drugs have been investigated in diseased patients [12]. There are a large number of papers to report global trends of the relation between the concentration of neuromodulators in the brain and behavioral observations among humans [28]. However, a quantitative evaluation of the performance of advanced functioning (e.g., attention, arousal) is very difficult, and moreover, difficult to find certain correspondences between the concentration of neuromodulators in the brain and the pathological symptoms of mental diseased patients.

In contrast, it is possible to make arbitrary designed experiments in animals, e.g. rodents [29], monkeys with use of genetical technology and so on [30] but in these cases, quantitative analysis of behaviors is quite difficult also [31].

Therefore, we have to recognize that many unknown mechanisms still remain hidden behind the deep complexity of brain systems. In these situations surrounded by high hurdle, a candidate to overcome difficulty is

‘functional chaos in systems with large but finite degrees of freedom’, in which there is high degree of redundancy in information processing and/or executing advanced functions from microscopic scale such as biological macro molecules to macroscopic scale, e.g. neural network in the brain. Several examples are shown in the authors’ paper [32–36], where the basic idea may be extended to even mesoscopic and microscopic scale such like the phenomena introduced in the present paper.

Acknowledgements The contribution of S.N. to this work was partly supported by JSPS KAKENHI Grant Numbers JP20H04246, JP20K11985, JP20K12002, and the Cooperative Research Program of “Network Joint Research Center for Materials and Devices”.

Funding Open access funding provided by Okayama University.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. Koukouli F, Changeux JP (2020) Do nicotinic receptors modulate high-order cognitive processing? *Trends Neurosci* 43:550–564
2. Poorthuis RB, Mansvelder HD (2013) Nicotinic acetylcholine receptors controlling attention: behavior, circuits and sensitivity to disruption by nicotine. *Biochem Pharmacol* 86:1089–1098
3. Perry E, Walker M, Grace J, Perry R (1999) Acetylcholine in mind: a neurotransmitter correlate of consciousness? *Trends Neurosci* 22:273–280
4. (Editors) Akaike A, Shimohama S, Mitsu Y (2018) *Nicotinic Acetylcholine Receptor Signaling in Neuroprotection*, Open access book of Springer, <https://doi.org/10.1007/978-981-10-8488-1>
5. Wang J, Lindstrom J (2018) REVIEW ARTICLE: Orthosteric and allosteric potentiation of heteromeric neuronal nicotinic acetylcholine receptors. *Br J Pharmacol* 175:1805–1821
6. Haga T (2013) Molecular properties of muscarinic acetylcholine receptors. *Proceed Japan Acad, Series B* 89:226–256. <https://doi.org/10.2183/pjab.89.226>
7. Cheng X, Ivanov I, Wang H, Sine SM, McCammon JA (2007) Nanosecond-timescale conformational dynamics of the human $\alpha 7$ nicotinic acetylcholine receptor. *Biophys J* 93:2622–2634
8. Unwin N (2005) Refined structure of the nicotinic acetylcholine receptor at 4 Å resolution. *J Mol Biol* 346:967–989
9. Miyazawa A, Fujiyoshi Y, Unwin N (2003) Structure and gating mechanism of the acetylcholine receptor pore. *Nature* 423:949–955
10. Changeux JP (1993) Chemical signaling in the brain. *Scient Am* 269:58–62

11. Bocquet N, Nury H, Baaden M, Poupon CL, Changeux JP, Delarue M, Corringer PJ (2009) X-ray structure of a pentameric ligand-gated ion channel in an apparently open conformation. *Nature* 457:111–114
12. Hurst R, Rollema H, Bertrand D (2013) Nicotinic acetylcholine receptors: from basic science to therapeutics. *Pharmacol Therapeut* 137:22–54
13. Mitra A, Bailey TD, Auerbach AL (2004) Structural dynamics of the M4 transmembrane segment during acetylcholine receptor gating. *Structure* 12:1909–1918
14. Changeux JP, Edelman SJ (2005) Nicotinic acetylcholine receptors. Odile Jacob Publishing Corporation, New York
15. Millar NS, Gotti C (2009) Diversity of vertebrate nicotinic acetylcholine receptors. *Neuropharmacology* 56:237–246
16. Zhao Y, Liu S, Zhou Y, Zhang M, Chen H, Xu HE, Sun D, Liu L, Tian C (2021) Structural basis of human $\alpha 7$ nicotinic acetylcholine receptor activation. *Cell Res* 31:713–716
17. Lu W, Liu Z, Fan X, Zhang X, Qiao X, Huang J (2022) Nicotinic acetylcholine receptor modulator insecticides act on diverse receptor subtypes with distinct subunit compositions. *PLoS Genet* 18:e1009920
18. Unwin N, Fujiyoshi Y (2012) Gating movement of acetylcholine receptor caught by plunge-freezing. *J Mol Biol* 422:617–634
19. Evans D, Cohen EGD, Morriss GP (1993) Probability of second law violations in shearing steady states. *Phys Rev Lett* 71:2402–2404
20. Wonnacott S, Isabel B, Millar NS, Tzartos SJ (2018) Themed section: nicotinic acetylcholine receptors. *Br J Pharmacol* 175:1783–2129
21. Esmaelia S, Murphy K, Swords GM, Ibrahima BA, Brownb JW, Llano DA (2019) Visual hallucinations, thalamocortical physiology and Lewy body disease: a review. *Neurosci Biobeh Rev* 103:337–351
22. Eversfield CL, Orton LD (2019) Auditory and visual hallucination prevalence in Parkinson's disease and dementia with Lewy bodies: a systematic review and meta-analysis. *Psychol Med* 49:2342–2353
23. Poorthuis RB, Bloem B, Schak B, Wester J, de Kock CP, Mansvelder HD (2013) Layer-specific modulation of the prefrontal cortex by nicotinic acetylcholine receptors. *Cereb. Cortex* 23:148–161
24. Cotton J, Edwards J, Rahman MA, Brumby S (2018) Cholinesterase research outreach project (CROP): point of care cholinesterase measurement in an Australian agricultural community. *Environ Health* 17:31
25. Hershey LA, Coleman-Jackson R (2019) Pharmacological management of dementia with lewy bodies. *Drugs Age* 36:309–319
26. Haken H (1996) Principles of brain functioning. Springer, Berlin & Heiderberg
27. Freeman WJ (1997) Societies of brains. Taylor & Francis, New York
28. Lewis AS, Picciotto MR (2020) Regulation of aggressive behaviors by nicotinic acetylcholine receptors: animal models, human genetics, and clinical studies. *Neuropharmacology* 167:107929
29. Terry AV Jr, Callahan PM (2019) Nicotinic acetylcholine receptor ligands, cognitive function, and preclinical approaches to drug discovery. *Nicot Tobacco Res* 21:383–394
30. Changeux JP (2010) Nicotine addiction and nicotinic receptors: lessons from genetically modified mice. *Nat Rev Neurosci* 11:389–401
31. St Peters M, Demeter E, Lustig C, Bruno JP, Sarter M (2011) Enhanced control of attention by stimulating mesolimbic-cortical cholinergic circuitry. *J Neurosci* 31(26):9760–9771
32. Nara S, Fujii H, Tsukada H, Tsuda I (2022) Visual hallucinations in dementia with lewy bodies originate from necrosis of characteristic neurons and connections in three-module perception model. To be published recently in *Scientific Reports*
33. Yano T, Goto Y, Nagaya T, Tsuda I, Nara S (2017) Pseudo-neuron device and firing dynamics of their networks similar to neural synchronizing phenomena between far local fields in brain. *Advances in cognitive neurodynamics*, Vol 6, In: Proceedings of the 6th international conference on cognitive neurodynamics, pp 109–118
34. Nara S, Soma K, Yamaguti Y, Tsuda I (2020) Constrained chaos in three-module neural network enables to execute multiple tasks simultaneously. *Neurosci Res* 156:217–224
35. Tsuda I (2001) Toward an interpretation of dynamic neural activity in terms of chaotic dynamical systems. *Behav Brain Sci* 24:793–810
36. Yamaguti Y, Tsuda I (2021) Functional differentiations in evolutionary reservoir computing networks. *Chaos* 31:013137

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.